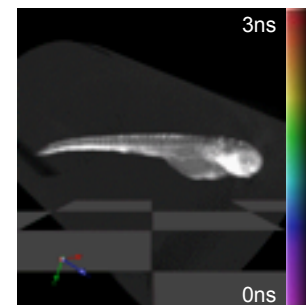
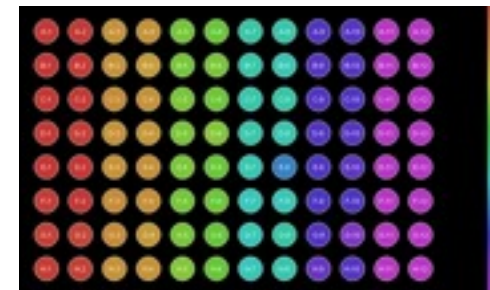
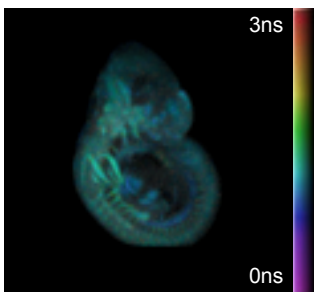
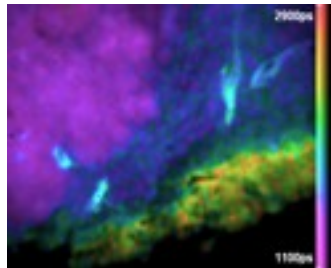
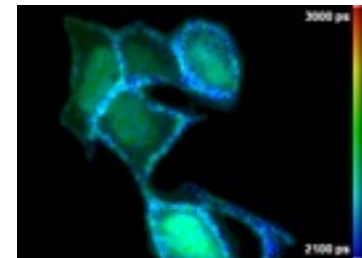
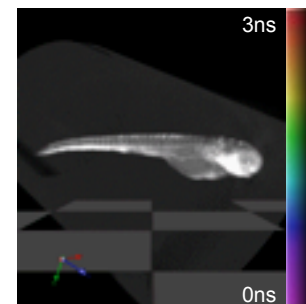
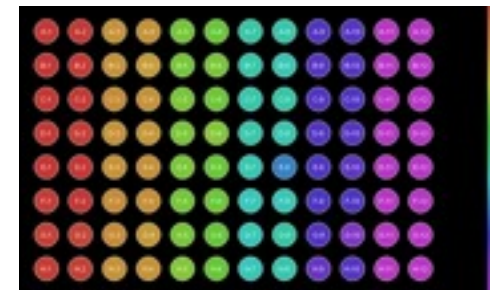
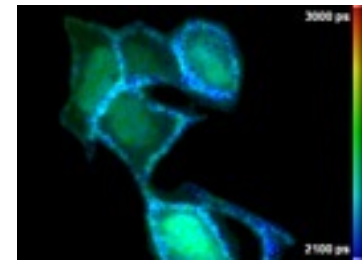


Fluorescence Lifetime Imaging (FLIM) and OMERO.

Yuriy Alexandrov
Sean Warren
Ian Munro
Chris Dunsby
Paul French

Photonics Group,
Physics Department
Imperial College London





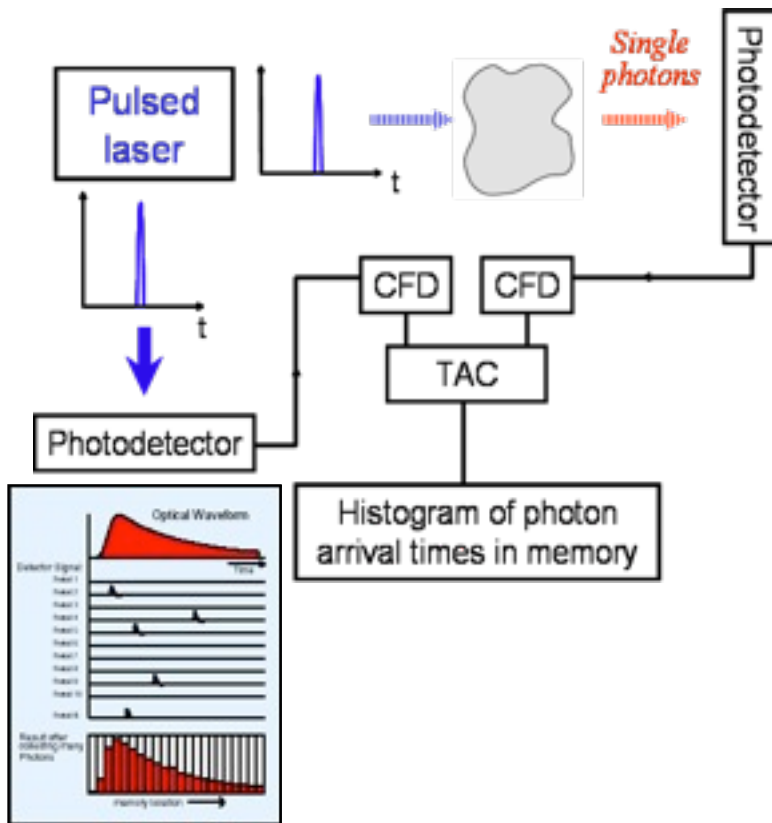
- **Introduction to our data**

- Time-Domain FLIM (TCSPC & time-gated)
- Förster Resonant Energy Transfer (FRET)
- *Multi-dimensional data (x y Z C T relative-time)*
- *Time-lapse, multiwell plate, z-stack*

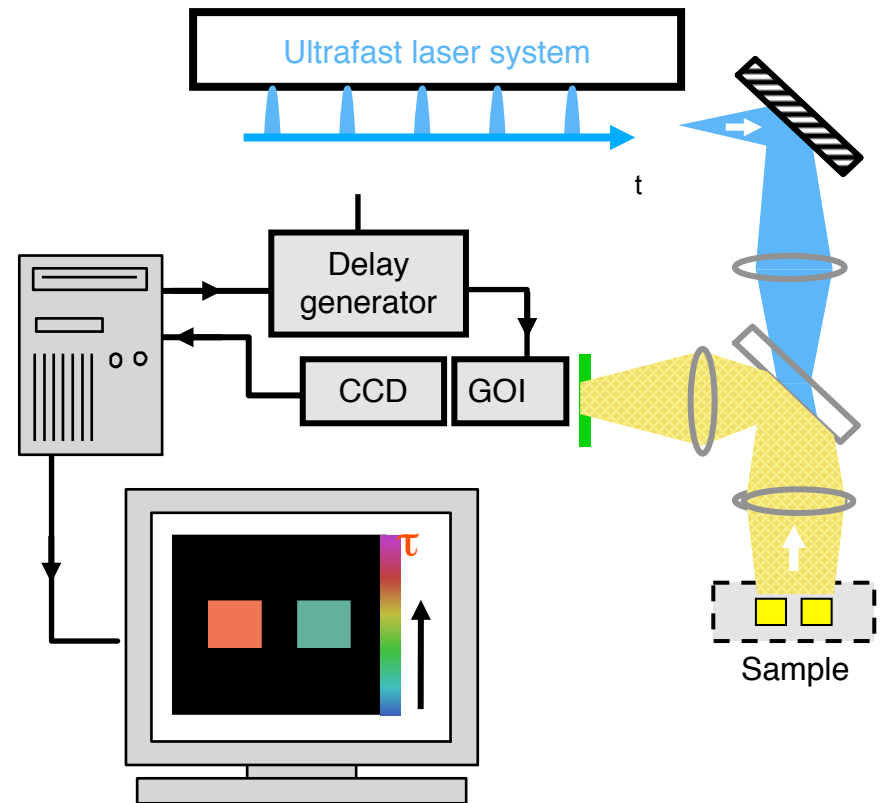
- **FLIMfit**

- *Open source tool to Fit TCSPC & time-gated FLIM data*
- *Integrated with OMERO*

(Time domain) FLIM technology

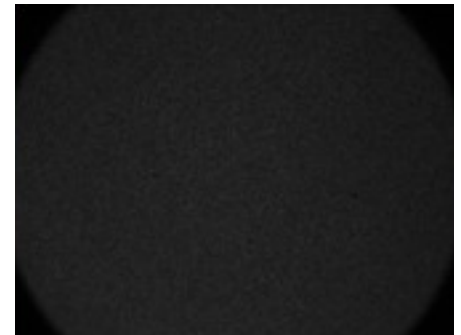
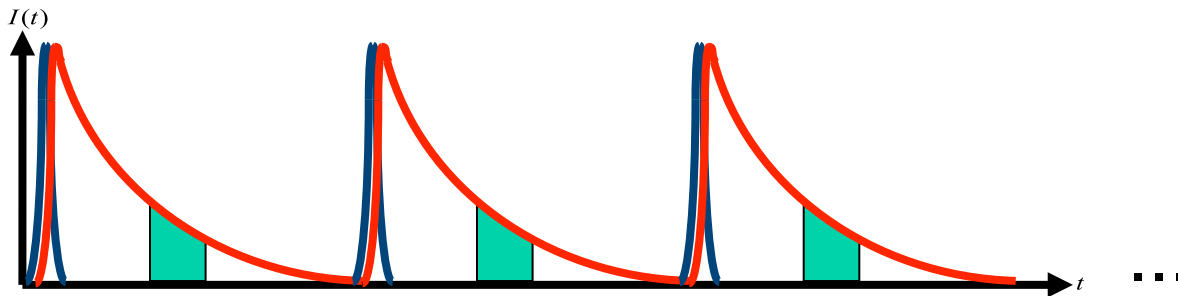
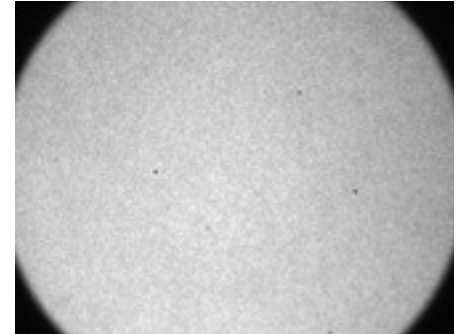
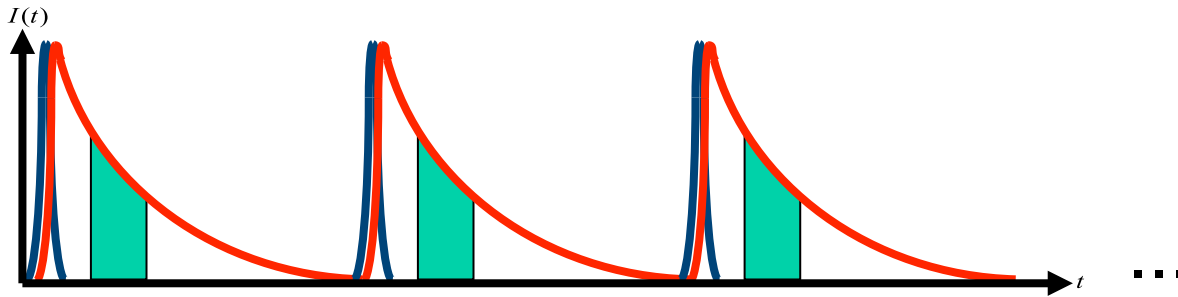


Confocal TCSPC



Wide-field time-gated imaging

Time-gated FLIM: Gated optical intensifier

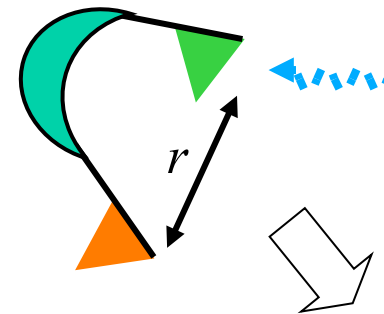
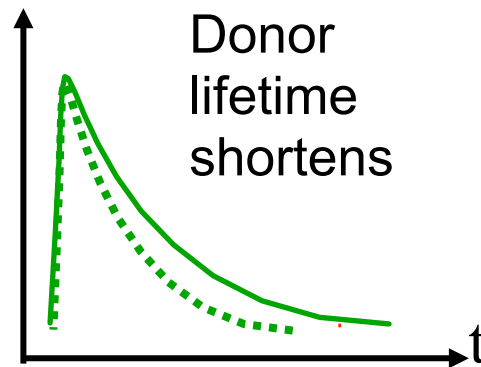
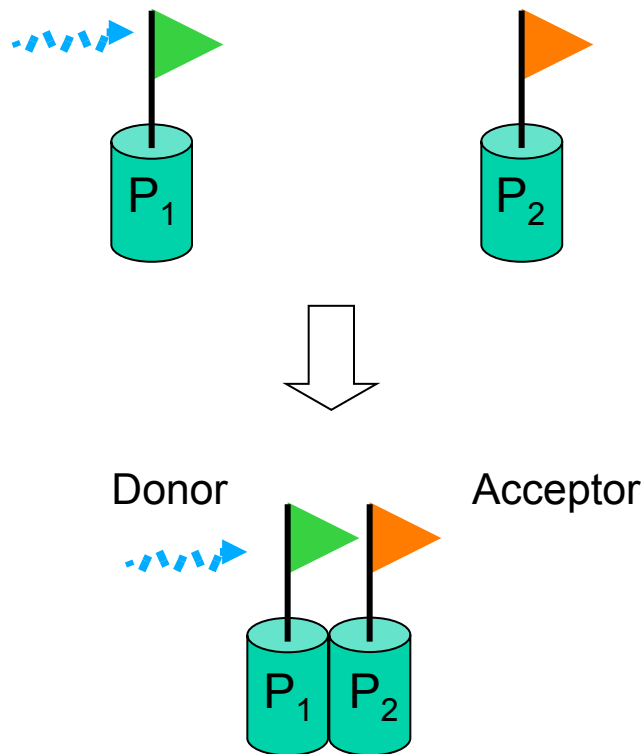


FLIM FRET to read out interactions & dynamics

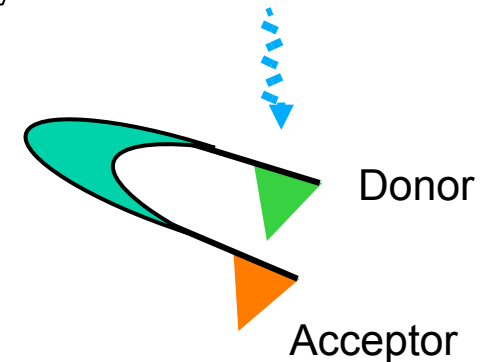
Förster Resonant Energy Transfer between fluorescent molecules over short (< 20 nm) distances : *dipole-dipole interaction (NO PHOTONS)*

e.g. protein binding

e.g. change in conformation

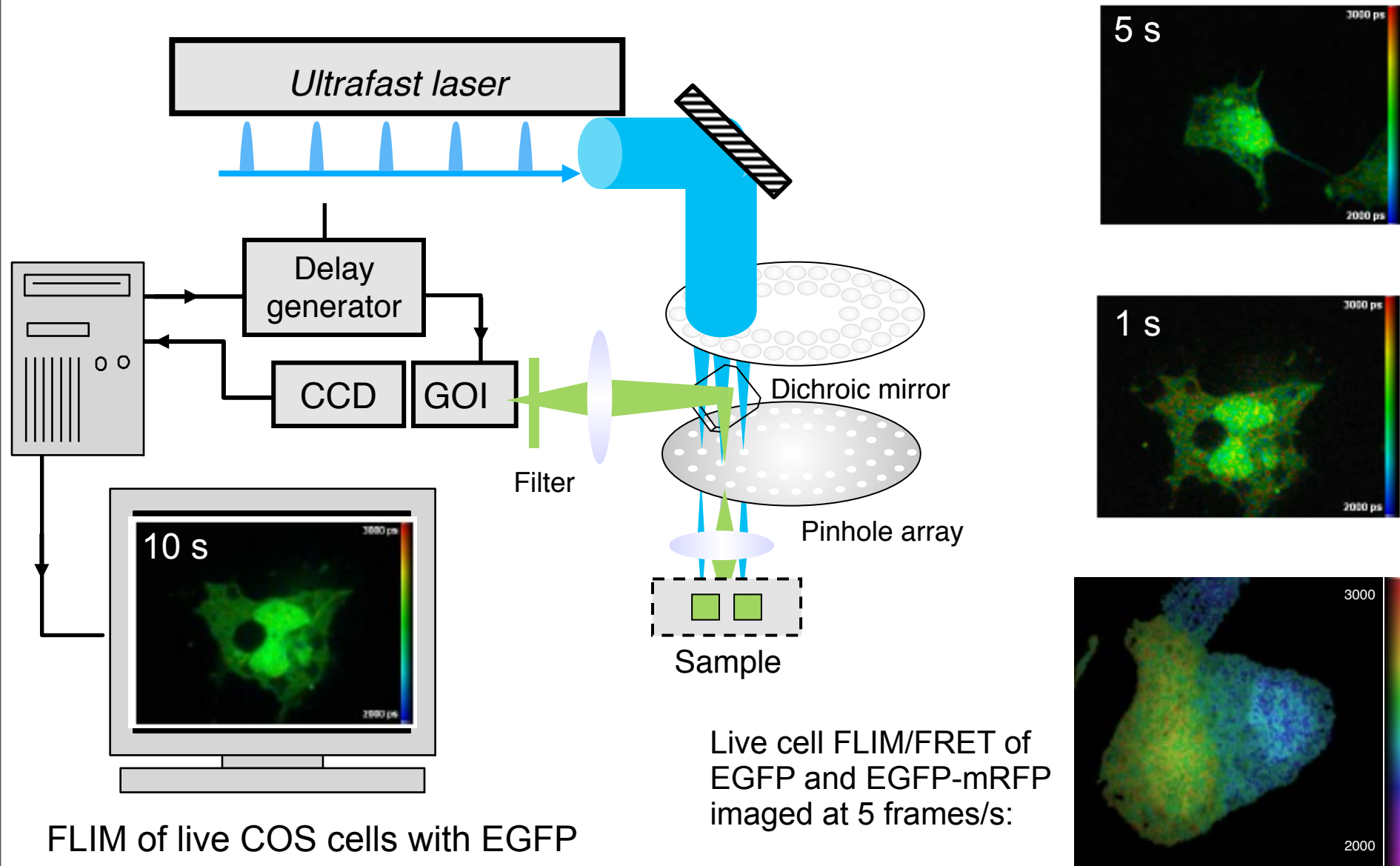


$$r = R_0 \left(\frac{1}{\eta_{FRET}} - 1 \right)^{1/6}$$

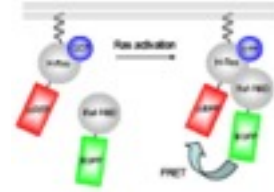


$$I = I_0 \left(\alpha e^{-t/\tau_{DA}} + (1-\alpha) e^{-t/\tau_D} \right) \quad \eta_{FRET} = 1 - \tau_{DA} / \tau_D$$

Wide-field optically-sectioned FLIM

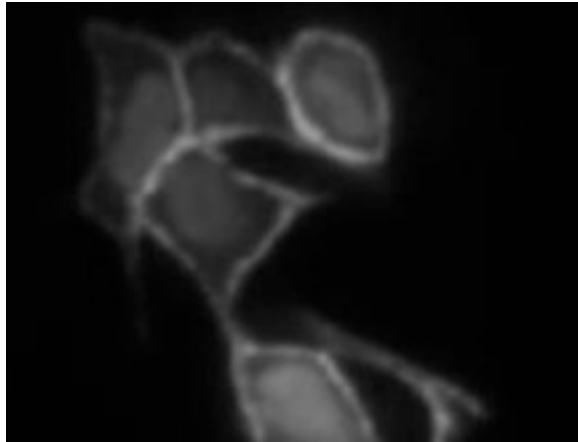


Nipkow FLIM-FRET of Raf RBD/Ras-mRFP

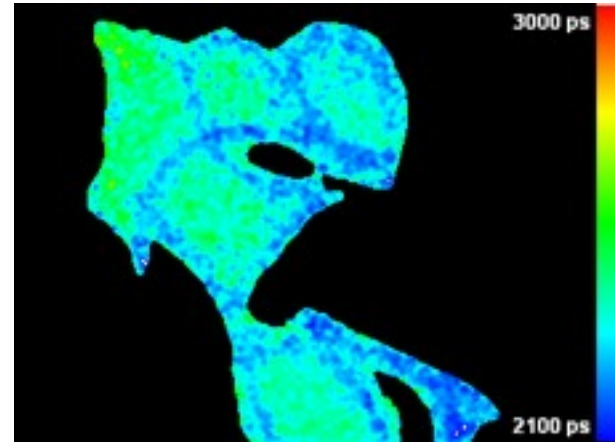


Imaged after 10 minutes stimulation with EGF (5 s FLIM acquisition)

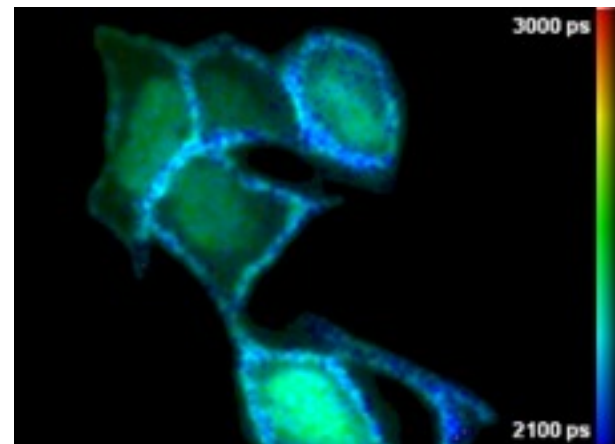
Raf-RBD-EGFP



Intensity image



FLIM of Raf-RBD-EGFP



(RBD = Ras Binding Domain)

Grant et al. Opt. Exp. (2007)

Time-lapse FLIM-FRET of $[IP_3]$ in MEFs experiencing a PDGF gradient

MEF expressing mTqFP-YFP “LIBRA” biosensor

⇒ IP_3 oscillations observed

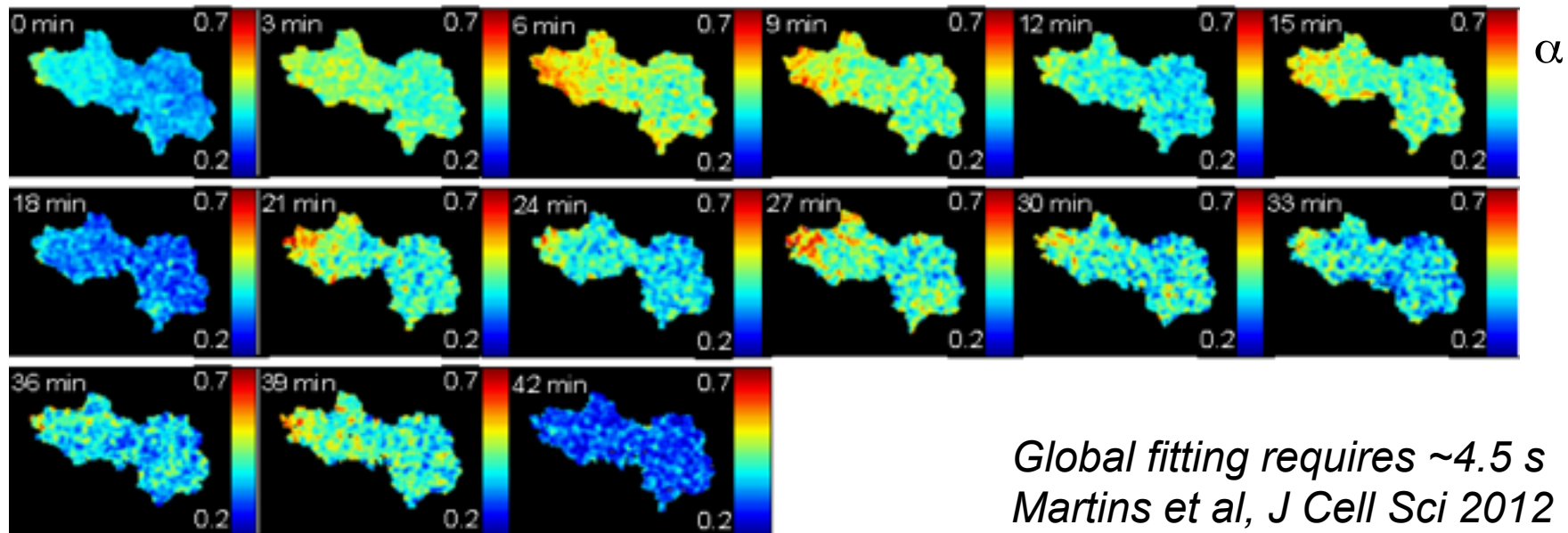
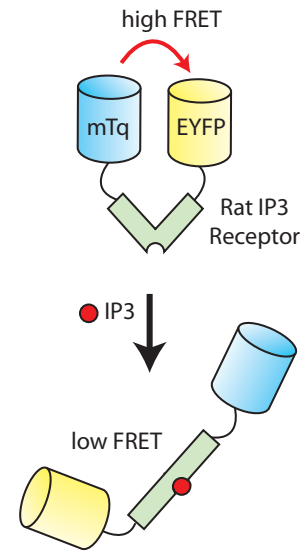
⇒ higher IP_3 concentration in direction of stimulation

Use global fitting across series to biexponential decay model

$$I_{GFP} = I_0 \left(\alpha e^{-t/\tau_1} + (1-\alpha) e^{-t/\tau_2} \right)$$

$$\tau_1 \text{ (low FRET)} = 3.2 \text{ ns}$$

$$\tau_2 \text{ (high FRET)} = 0.78 \text{ ns}$$



*Global fitting requires ~4.5 s
Martins et al, J Cell Sci 2012*

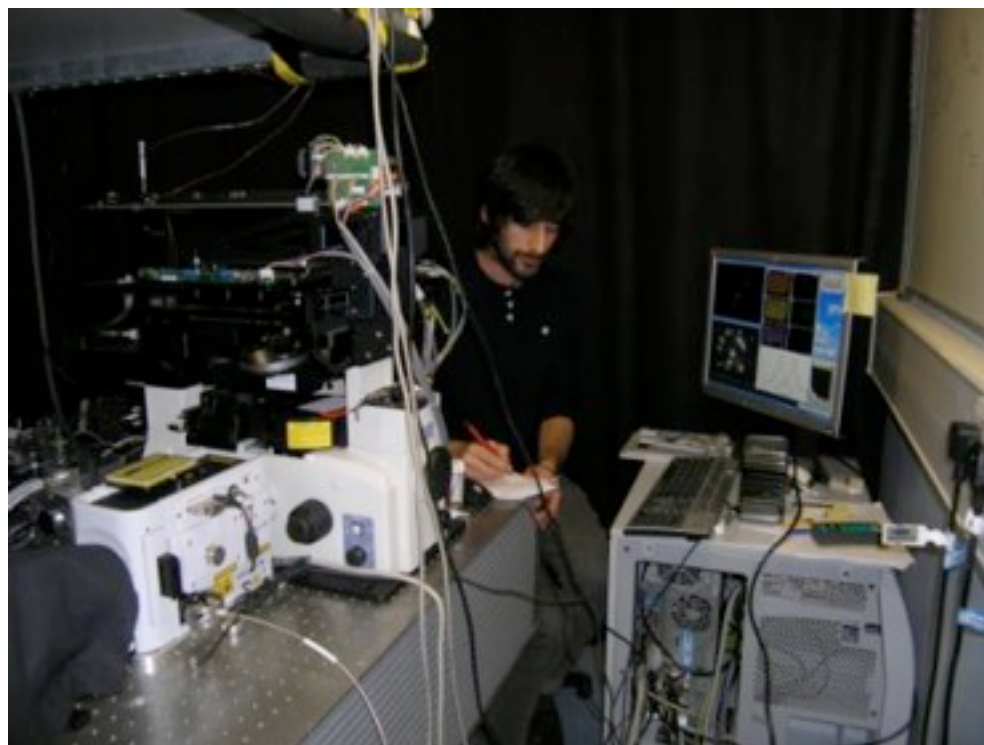
TSB project: prototype FLIM multiwell plate reader

(based on GE Healthcare IN Cell 1000)

Established wide-field
multiwell plate reader

- + *Yokogawa CSU-X (more efficient)*
- + *wide-field time-gating*
- + *supercontinuum excitation source*
- + *FLIM/segmentation analysis*
- + *prescan mode*

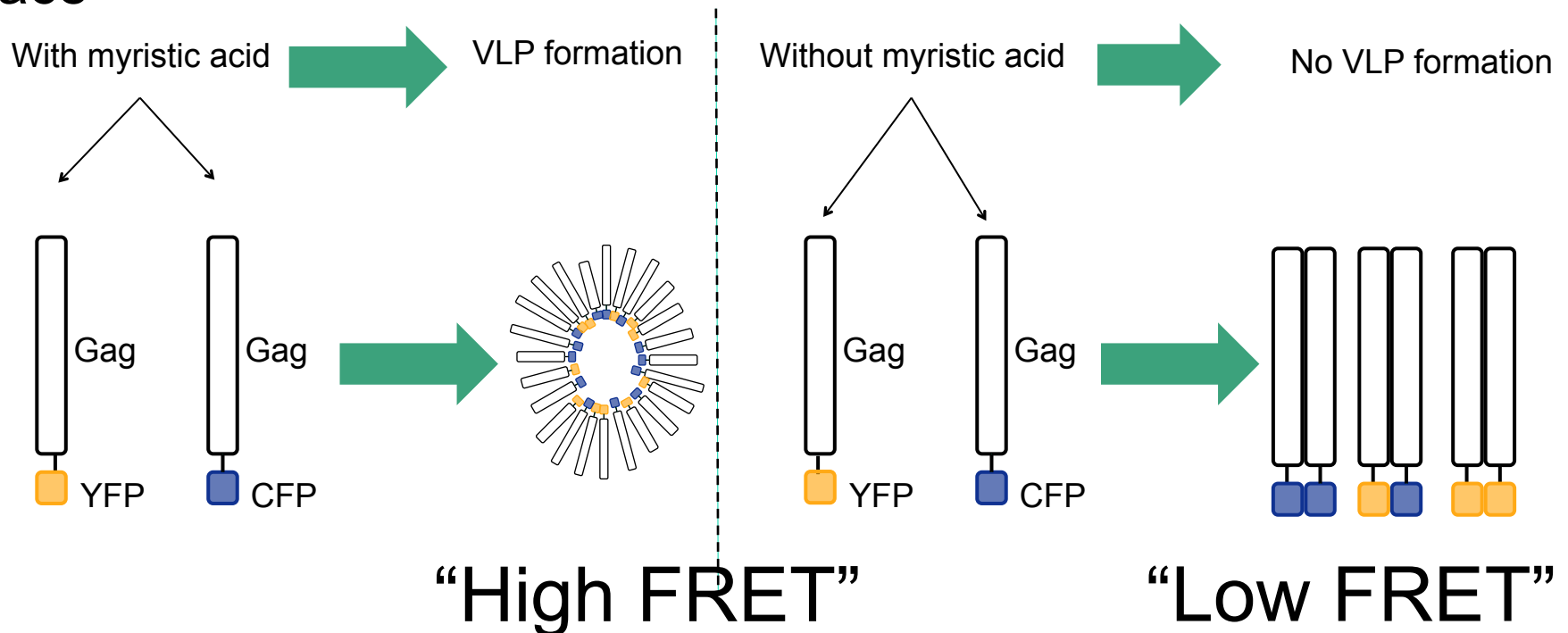
⇒ <~15 min/96 well plate for
FLIM of FP-labelled live cells

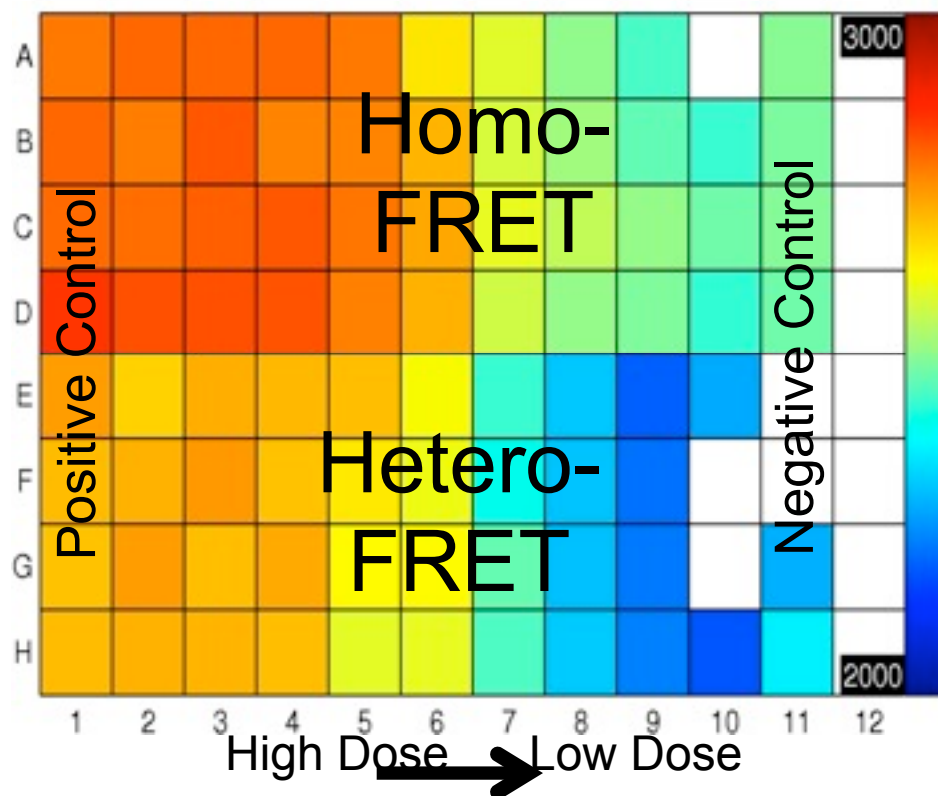


Kumar et al. ChemPhysChem (2011)

HIV-1 Gag Assay: Fluorescently labelled Gag protein

- HIV-1 Gag proteins are dependent on a 'myristic switch' mechanism for membrane binding
- Removal of the first N-terminal glycine residue removes the ability for myristoylation → no VLP formation should take place



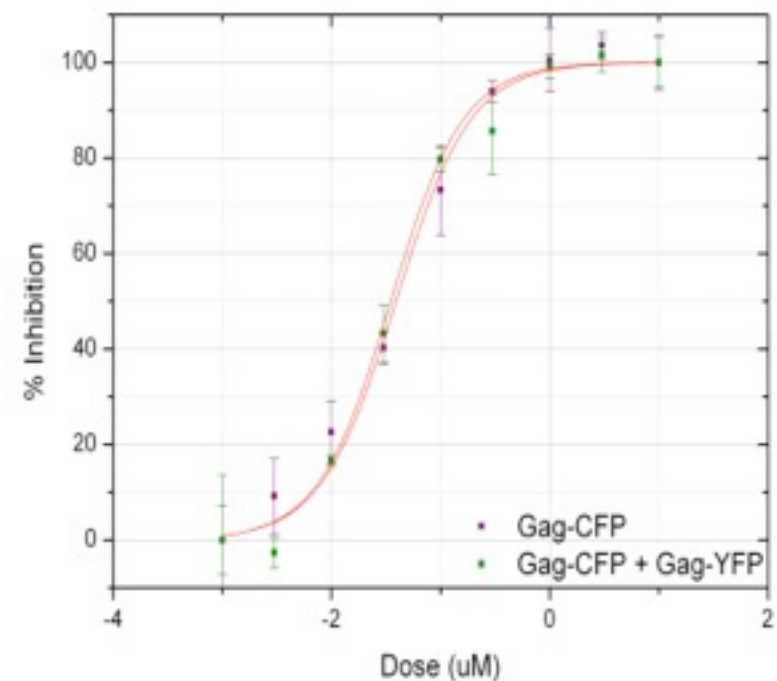
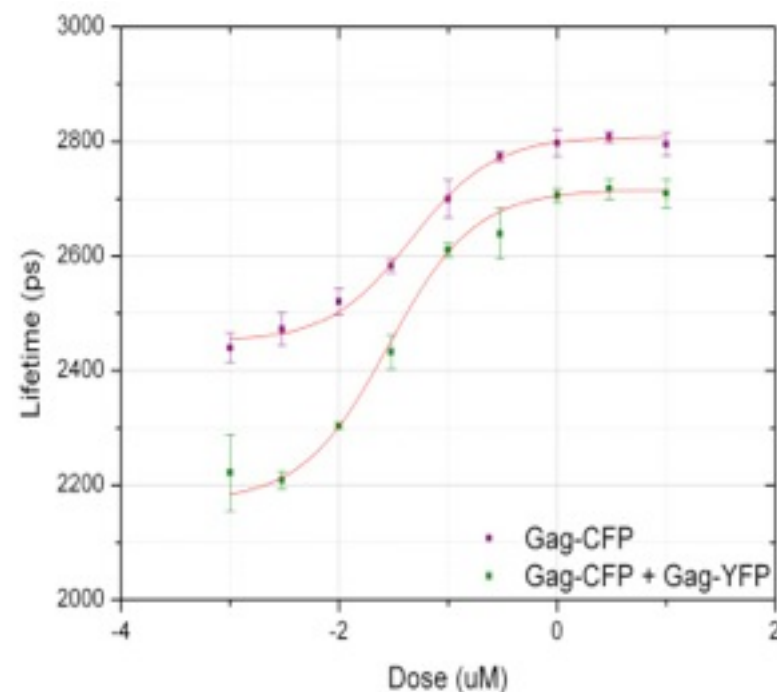


- Assay performance measured by Z' parameter:

$$Z' = 1 - \left(\frac{3 * [\sigma_{pos} + \sigma_{neg}]}{\mu_{pos} - \mu_{neg}} \right)$$

$$Z'_{\text{Homo-FRET}} = 0.63$$

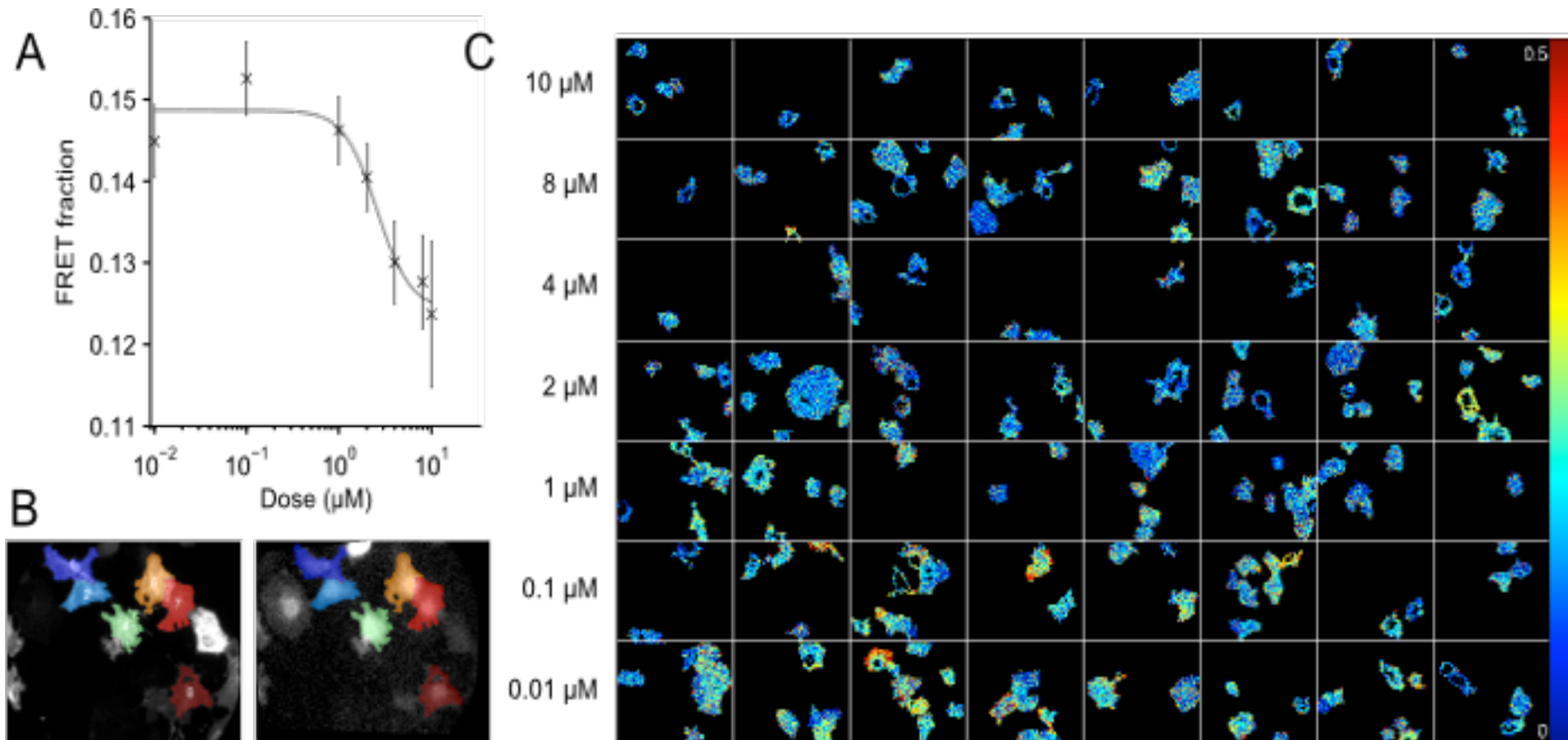
$$Z'_{\text{Hetero-FRET}} = 0.45$$



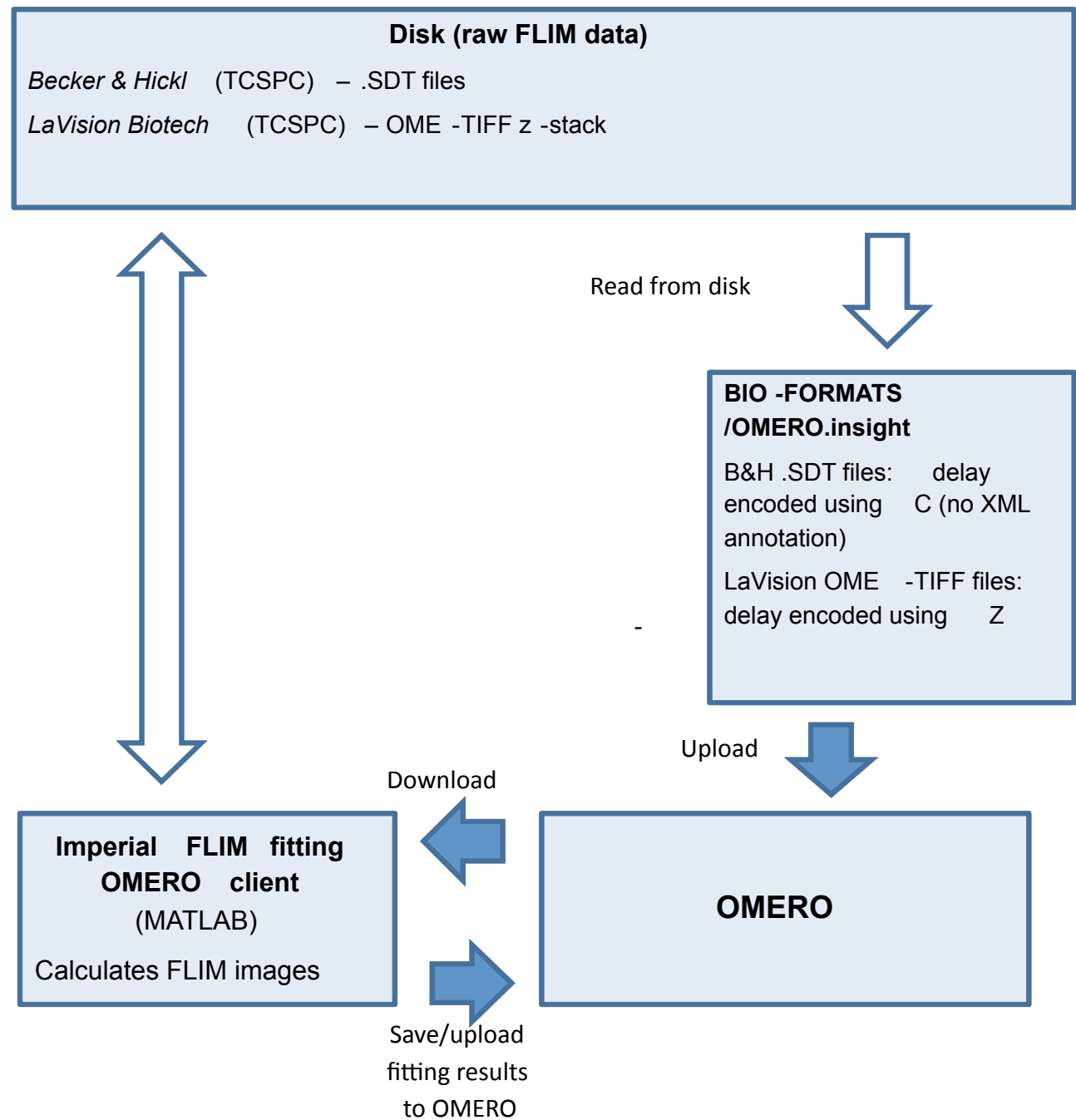
Global analysis of FLIM data

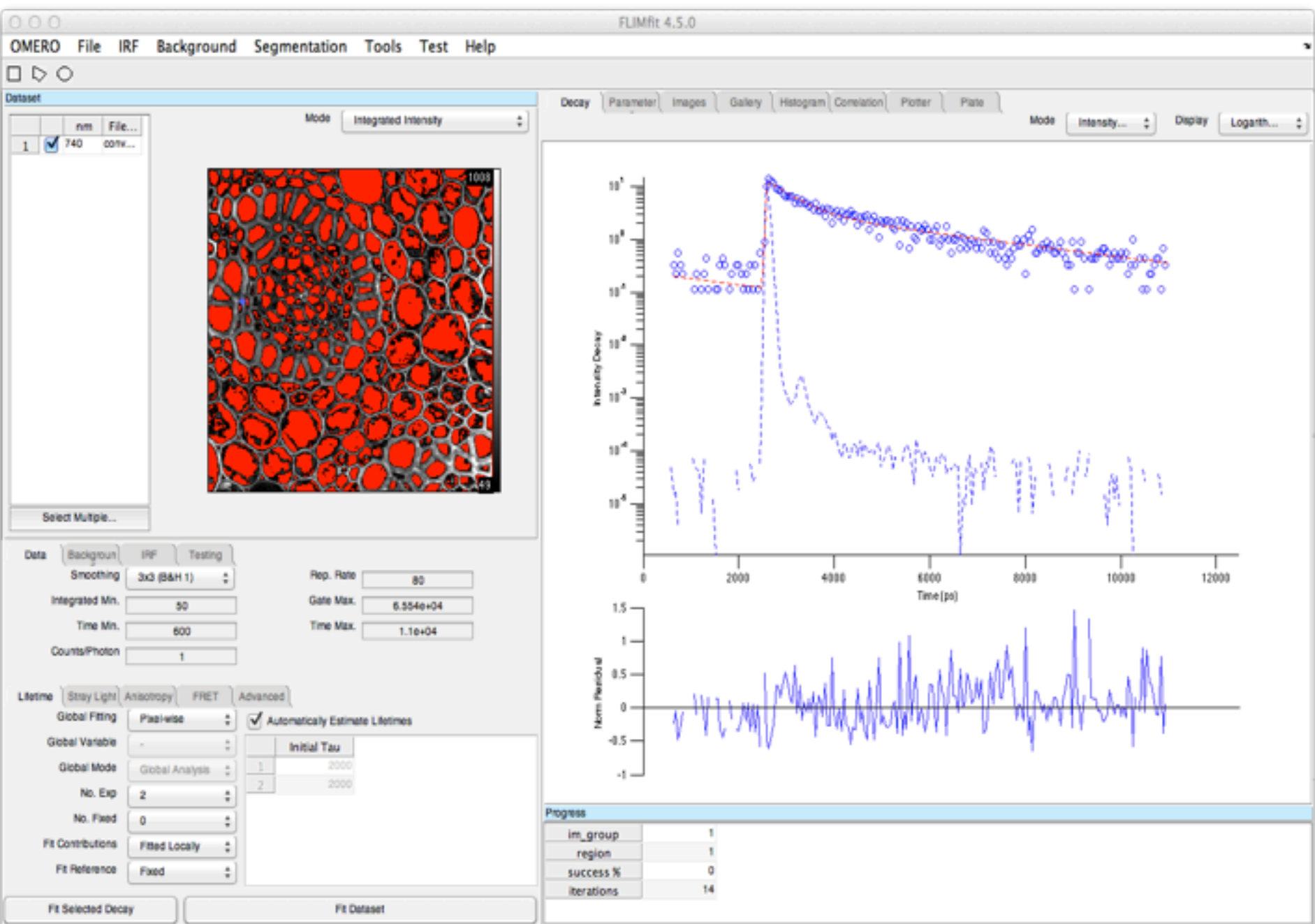
Global fitting enables complex decay models (e.g. for FRET) with modest (100's) photon numbers/pixel

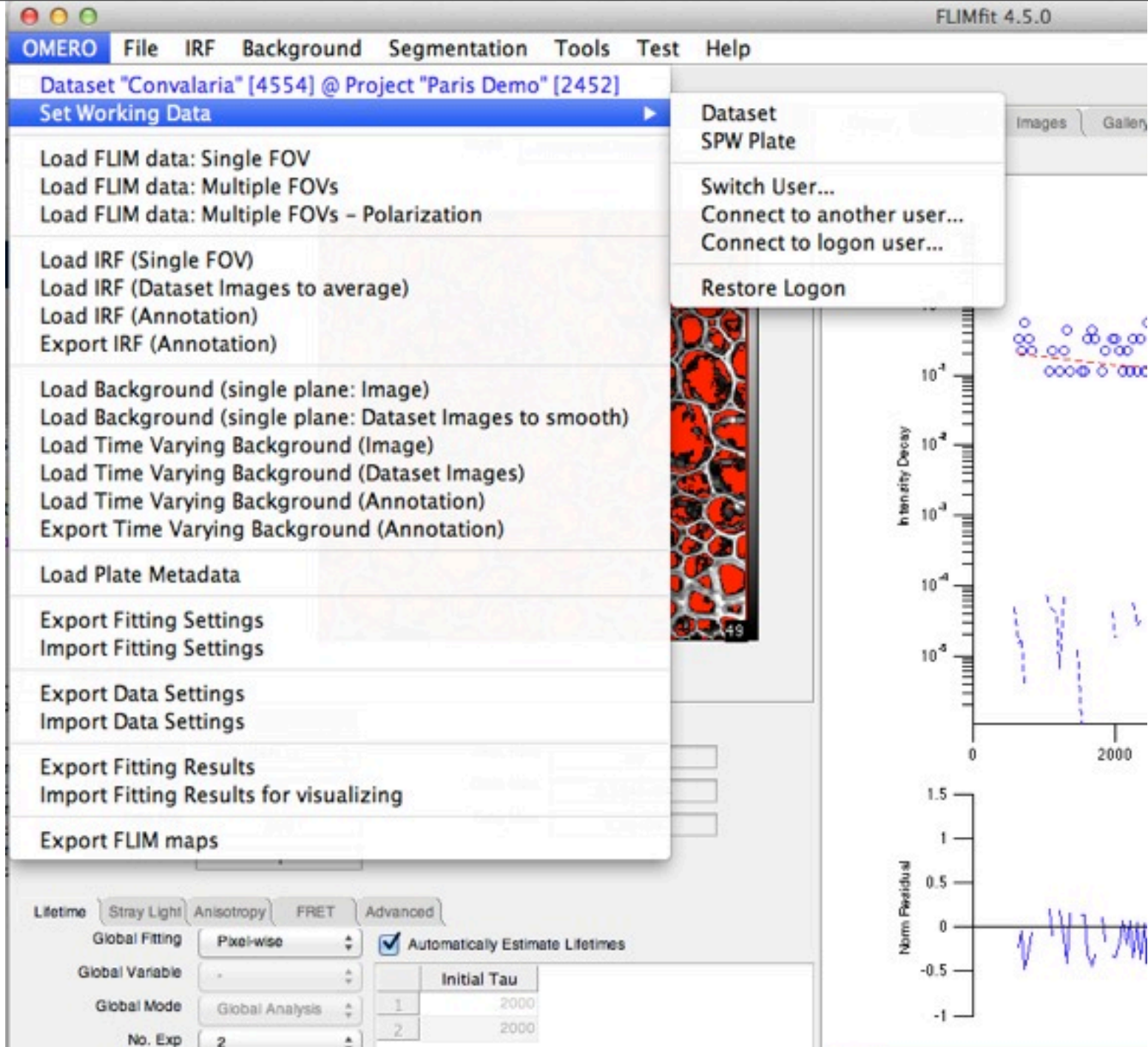
e.g. assay of inhibitor IPA-3 on interaction between Rac1 and p21-activated kinase read out with mTurquoise FLAIR biosensor in COS-7 cells stimulated with EGF

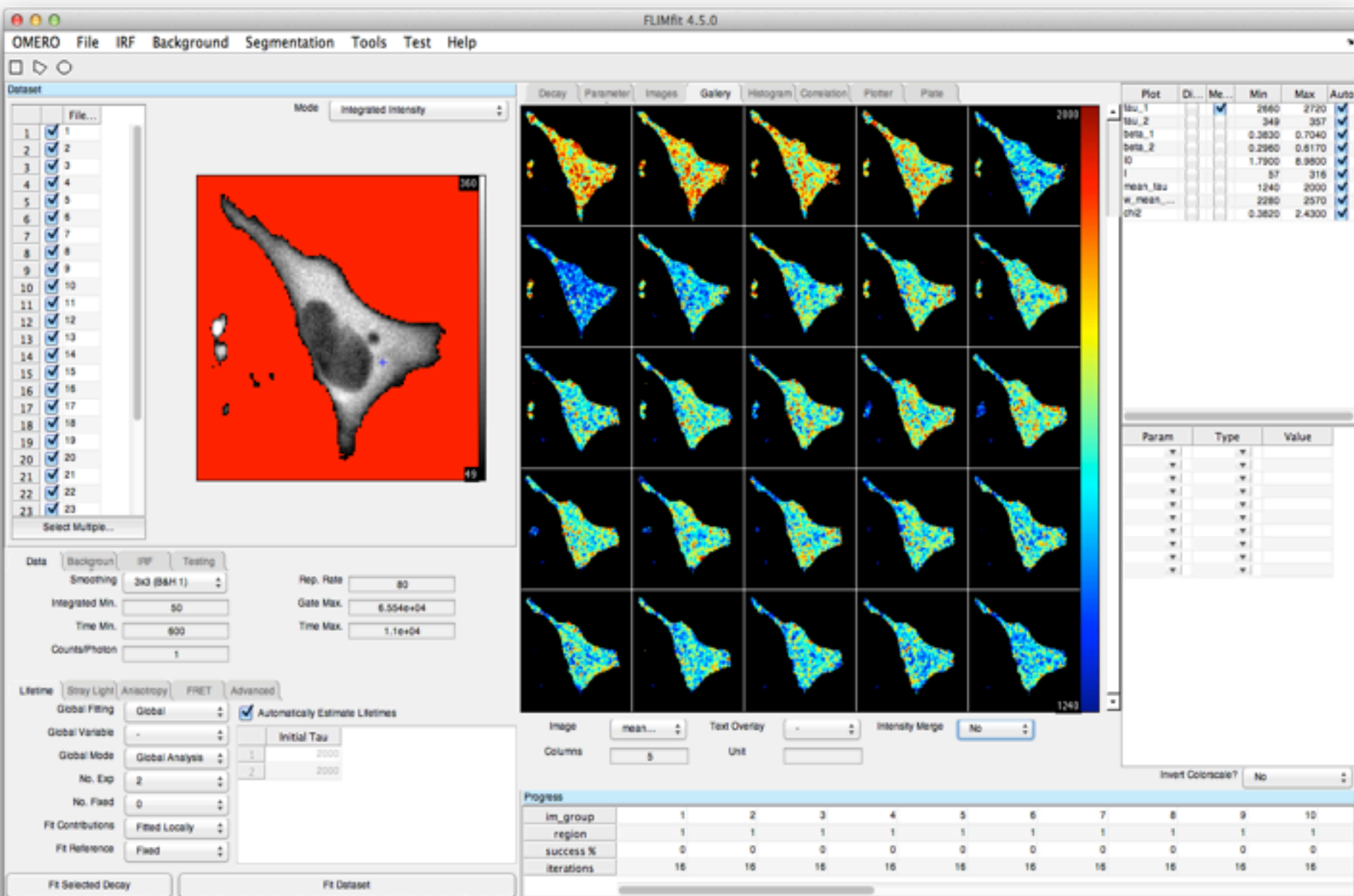


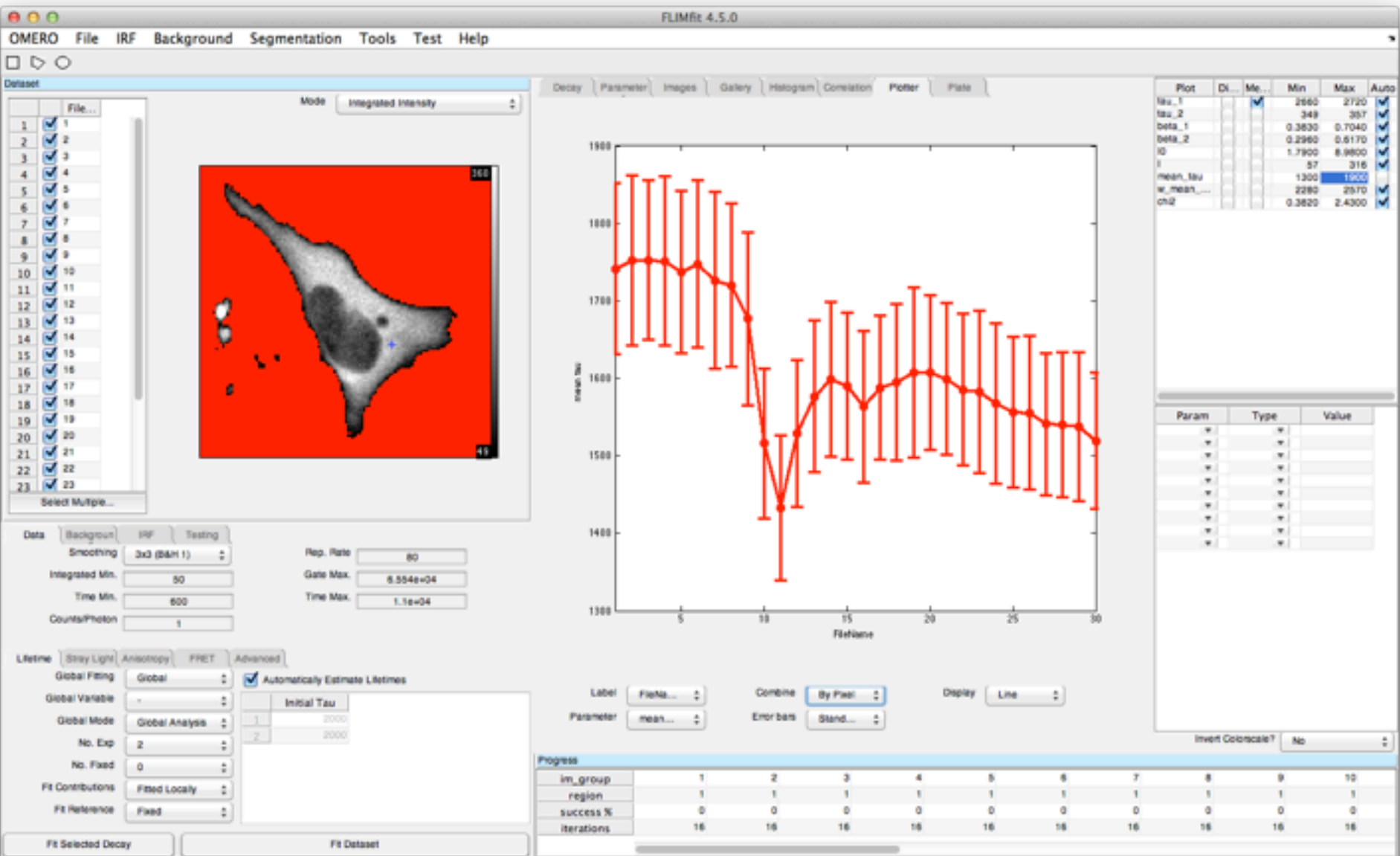
FLIMfit: an OMERO client in MATLAB/C++

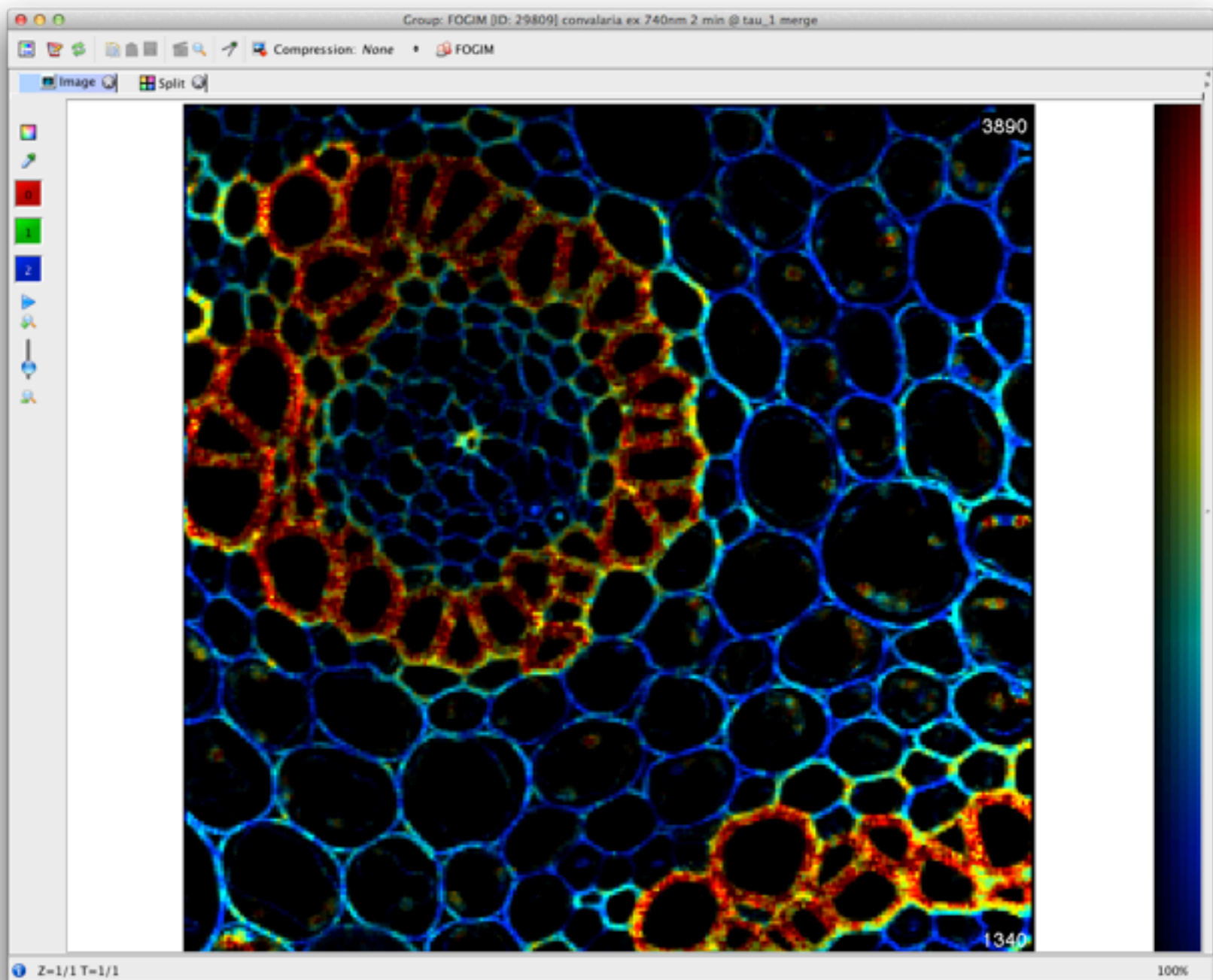












FLIMfit: an OMERO client

Sean Warren + ...

Open Source, cross-platform

Two time-domain FLIM modalities (TCSPC, Gated)

Multi-threaded

Global Analysis

Two forms of IRF (scatterer & reference fluorophore)

Support for simultaneous analysis of large datasets (100's images)

Supports time-varying background (TVB)

Maximum likelihood option

PLoS One : Rapid global fitting of large fluorescence lifetime
imaging microscopy datasets S.C. Warren, A. Margineanu, D.
Alibhai, D.J. Kelly, C. Talbot, Y. Alexandrov, I. Munro, M. Katan, C.
Dunsby*, P.M.W. French*

<http://www.openmicroscopy.org/site/products/partner/flimfit>

<https://github.com/openmicroscopy/Imperial-FLIMfit>

Multidimensional fluorescence imaging

Imperial College
London



Dominic Alibhai, Natalie Andrews, Lingling Chen, Sergio Coda, Pieter de Beule, David Grant, Douglas Kelly, Romain Laine, Hugh Manning, Dylan Owen, Stephane Oddos, Rakesh Patalay, Tom Robinson, Hugo Sinclair, Hugh Sparks, Sean Warren, Neil Galletly, Yuriy Alexandrov, Egidijus Aukorius, Alice Brown, Sunil Kumar, Peter Lanigan, Martin Lenz, Anca Margineanu, Ewan McGhee, Ian Munro, Jose Requejo-Isidro, Gordon Kennedy, Daniel Stuckey, Paul Tadrous, Harriet Taylor, Khadija Tahir, Clifford Talbot, James McGinty, Chris Dunsby, Mark Neil, Paul French

Imperial College London

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