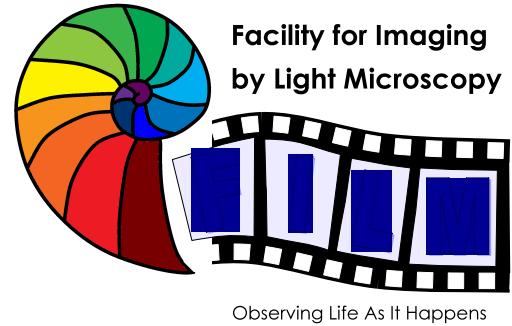


OMERO Implementation at Imperial College London

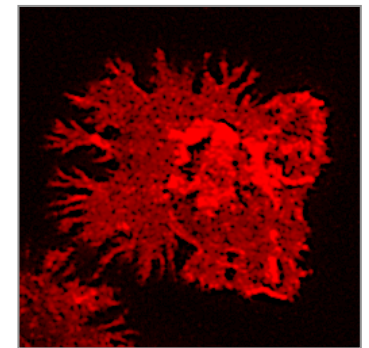
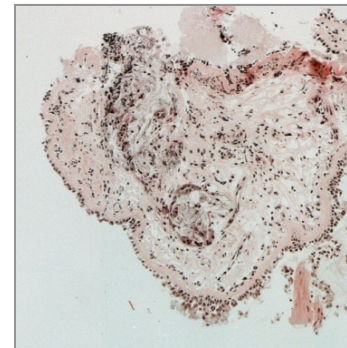
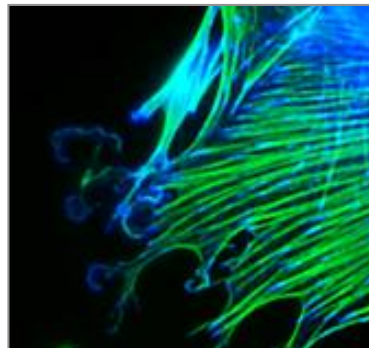
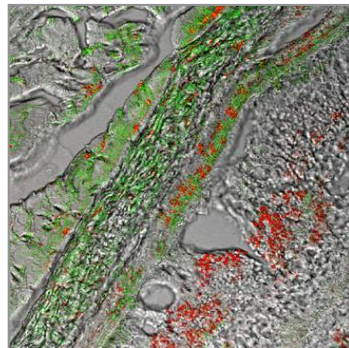
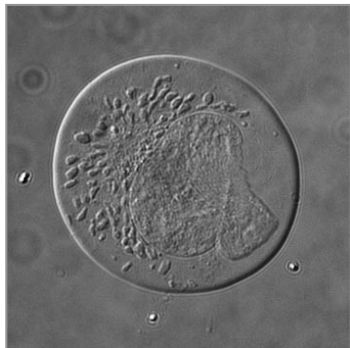
Imperial College
London



Bioinformatics
Support
Service

Martin Spitaler

Mark Woodbridge



OMERO Implementation at Imperial College London

- Installation, technical solutions, data integration: Bioinformatics Support Service (Mark Woodbridge, Chris Tomlinson)
- Testing & Users: Facility for Imaging by Light Microscopy (FILM) (Martin Spitaler, Christian Liebig, Steve Rothery), facility users, some non-facility users
- Time scale:
 - first test installation: April 2008
 - large-scale import test and feedback: June 2009
 - first tests with live data: December 2009
 - general roll-out to facility users: from January 2010

Deployment Strategy

- Extensive research
 - Hardware
 - Formats
- Working relationship
 - IT
 - Bioinformatics
 - Imaging
- Candidate Users/Projects
 - Scale
 - Formats
- Expansion
 - Infrastructure
 - Support

Deployment Issues

- Software upgrades
 - Planning and testing
 - Web Start
- Authentication
 - SSL
 - Directory
 - SSO
- Data in/out
 - Formats
 - Metadata
 - Archival (and integrity)
 - Access
- Onboarding
 - Scalability
 - Tutorials

Ongoing Work

- Data management
 - Experimental workflows
 - Long-term storage
- Software pipelines
- Server-side processing
- Institutional integration

OMERO/XperimentR (Assay)

The screenshot displays the Omero/XperimentR Assay interface. The main workspace shows a hierarchical workflow diagram:

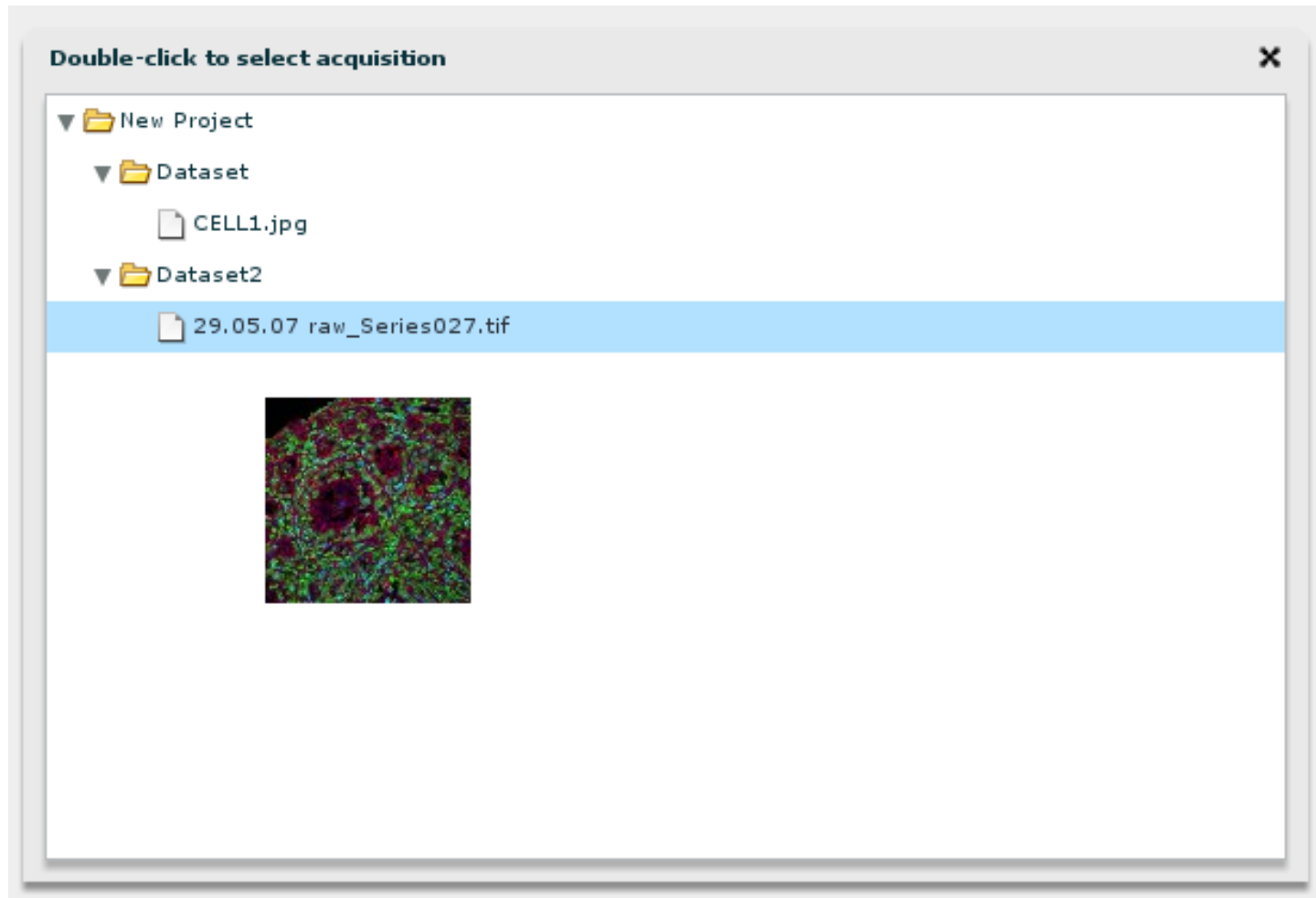
- Root node: A grey rounded square.
- Level 1: Three nodes in red boxes: a mouse icon, a mouse icon, and a mosquito icon.
- Level 2: Three nodes in green boxes: a flask with green liquid (connected to the first mouse icon), a test tube (connected to the second mouse icon), and a test tube (connected to the mosquito icon).
- Level 3: A microscope icon in a blue dashed box (connected to the flask).

At the bottom left of the workspace are navigation icons: a magnifying glass with a '1', a plus sign, a minus sign, and a hand icon. At the bottom right is an "Export study" button.

The right sidebar contains the "Assay Details" panel:

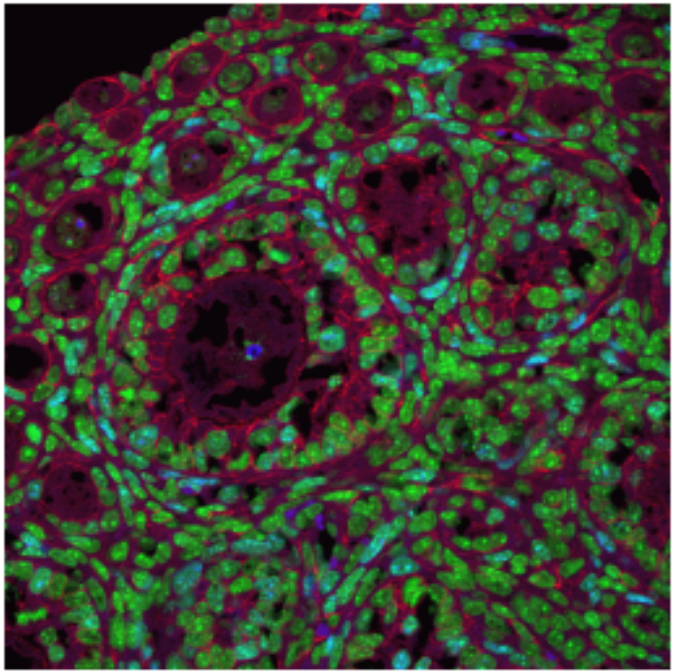
- Assay Details** (with a refresh icon)
- Date & Time assay performed:** 14/06/2010 (calendar icon) 11 (dropdown) hrs
- Assay Type:** Microscope
- Attach data** button
- Attach File** button
- Table with columns **File Name** and **Size (B)**
- Input fields: **Enter Term name** and **Enter Term**

OMERO/XperimentR (Repository)



OMERO/XperimentR (Image)

Click image to view in OMER0.web X



Assay Details

Date & Time assay performed:
14/06/2010 11 hrs 5 mins

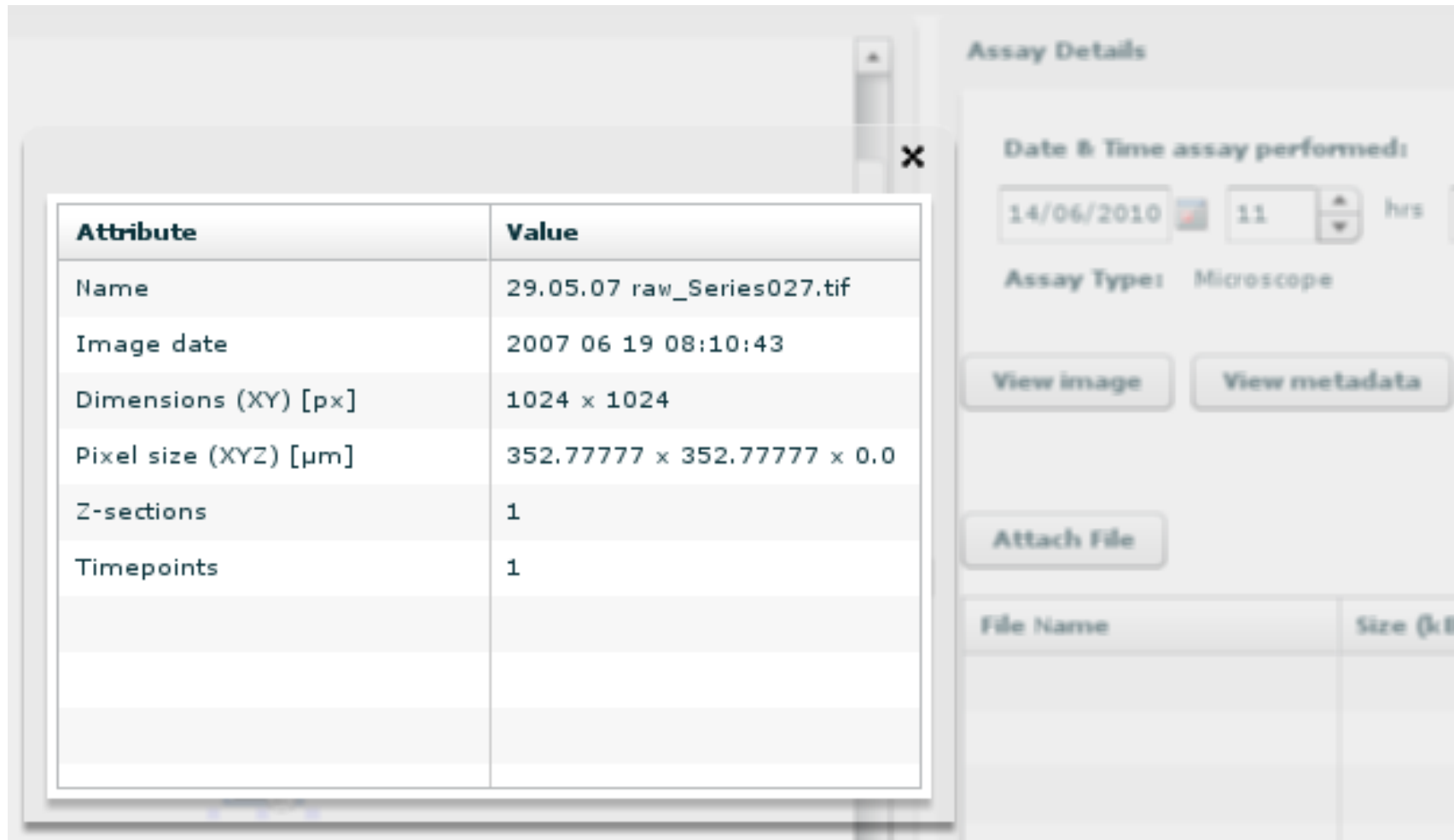
Assay Type: Microscope

View image View metadata Remove

Attach File

File Name	Size (kB)

OMERO/XperimentR (Metadata)



The screenshot displays the Omero XperimentR metadata interface. A modal window in the foreground shows a table of metadata attributes for a specific image. The background shows the 'Assay Details' section of the application.

Attribute	Value
Name	29.05.07 raw_Series027.tif
Image date	2007 06 19 08:10:43
Dimensions (XY) [px]	1024 x 1024
Pixel size (XYZ) [μm]	352.77777 x 352.77777 x 0.0
Z-sections	1
Timepoints	1

Assay Details

Date & Time assay performed: 14/06/2010 11 hrs

Assay Type: Microscope

View image View metadata

Attach File

File Name	Size (kB)

User overview

- Omero users:
 - FILM:
 - facility staff: microscope tests, user projects
 - researchers currently using OMERO: ~15
 - (potential users: ~380 individual microscopists / ~120 groups)
 - outside FILM:
 - EM (OMERO trial)
 - Photonics (fast Flim, OMERO trial)
- research areas: cell biology, developmental biology, immunology, infection biology, physiology, stem cell research, biological engineering
- methodologies: widefield, confocal, two-photon (in vivo), Flim, TIRF, PALM / STORM (in development), high-throughput screening

Microscope overview, file formats

- Widefield:
 - Zeiss Axiovert 200M with **Hamamatsu HClmage (*.TIFF, *.CXD)** and **Perkin Elmer Volocity (Volocity database format)**
- Confocal:
 - Zeiss LSM-510 (*.LSM)
 - Leica SP-2 (TIFF, *.LEI)
 - Leica-SP5 (LIF)
- Flim:
 - Becker-Hickel (*.SPC)
- HTS:
 - Cellomics (*.C01)

User experience - file formats

File type	Pixel data	Meta data
Hamamatsu HImage (*.TIFF, *.CXD)	yes	partially
Perkin Elmer Volocity (Volocity database format)	no	no
Zeiss LSM	yes	yes
Leica SP2 LEI	yes (mostly)	most
Leica SP5 LIF	yes (mostly)	most
Becker-Hickl SPC	no	no
Cellomics	not tested	not tested

User experience - file formats

Remaining issues:

- **Hamamatsu HImage (*.TIFF, *.CXD):**
 - XYZT not supported
 - channel information, exposure time, gain, ... not detected (time course is detected)
 - metadata are imported in raw format, but not interpreted
- **Volocity:**
 - not supported, indirect via OME-TIFF not possible, apparently due to an export fault
- **Leica SP2 LEI:**
 - lambda-scan imported, but as a time series (metadata not correct)
 - import 'unstable'
- **Leica SP5 LIF:**
 - lambda-scan as for LEI
 - MP laser settings not detected
- **Becker-Hickl SPC:**
 - not supported

User experience - usability

- **the core functionality of OMERO is seen as extremely useful:**
 - central repository of data from various microscopes
 - easy visualisation
 - easy access from any location
 - sharing (but too limited for routine usage)
 - quick access to archived data and OME-TIFF
- **visualisation:**
 - generally good, many good features (split screen, copy / paste rendering settings, greyscale,
 - rendering limitations limit its usability (visualisation is not quantitative, due to autocontrast, positive and negative controls appear with equal brightness)
 - the problem of side-by-side visualisation of many images is unsolved:
 - zoom too small in thumbnail view
 - split / greyscale visualisation not available
- **data archiving:**
 - in principle great solution for central data archiving
 - main problem: unreliable archiving (easy to forget archive tick box, not option to set it by default)
 - following from this, data duplication (local storage is preferred)

Wish list

Metadata:

- support for missing formats
- confocals: pinhole size as Airy Units (in addition to micrometer)

Data storage:

- it should be possible to make archiving non-optional in the preferences (or by administrator)
- when deleting data, the original image data should be deleted (or quarantined)

Usability:

- rendering!
 - option to turn it off by default throughout
 - must accept values outside min / max pixel values
- UNDO functions
- user preferences:
 - screen layout
 - default rendering / visualisation (e.g. min / max, greyscale, split, ...)
 - data archiving on

Sharing:

- sharing via INSIGHT
- access to archived file via sharing and / or export as OME-TIFF
- hierarchy preserved in shared files (sharing of whole folder / experiments)

Future plans

Improving implementation:

- improved integration in lab workflow (annotation / sharing)
- final data storage solution

Improving implementation:

- improved integration in lab workflow (annotation / sharing)
- final data storage solution

Improving data quality:

- annotation with (compulsory?) experimental metadata on upload
- integration with other data (Systems Biology)

Improving usability:

- integration with image analysis (Definiens, CellProfiler):
 - software should read image data from OMERO, analyse them and write data back into OMERO, together with analysis protocols (logs) and results
 - raw data, analysis logs and results should be automatically linked in OMERO
 - ideally, all data should be visualised in easily accessible form (e.g. multiwell plate layout)

Cooperations - THANKS

- FILM staff: Christian Liebig (now Tübingen), Steve Rothery, Mark Scott
- Bioinformatics Support Service: Mark Woodbridge, Chris Tomlinson, Sarah Butcher
- OMERO team Dundee: Jason Swedlow, Will Moore, ...
- Our patient users, first and foremost Marek Cebecauer (Imperial College / University Prague)
- Hamamatsu support, Leica engineers, Definiens Support