

# Image content-based searching and tools for image-derived modeling

Robert F Murphy

Ray & Stephanie Lane Professor of Computational Biology and  
Professor of Biological Sciences, Biomedical Engineering and Machine Learning  
External Senior Fellow, Albert Ludwig University of Freiburg

RAY AND STEPHANIE LANE  
Center for Computational Biology

**Carnegie Mellon**

ALBERT-LUDWIGS-  
UNIVERSITÄT FREIBURG



**FRIAS**  
FREIBURG INSTITUTE  
FOR ADVANCED STUDIES

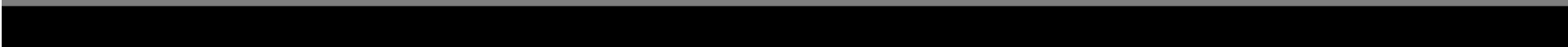
LIFE SCIENCES - LIFENET

# Content-based image search

- Also called Query-by-image-content (QBIC)
- Find images whose content, as reflected by image features, is similar to one or more query images
- Can use positive and/or negative examples
- Can be iterative (relevance feedback)



# Why content-based search?

- In compound screening, to see whether anyone else in the organization (or the world) has seen a particular pattern before
  - In proteomics, to find other proteins that show a subcellular location that is difficult to assign
- 



# OMERO.searcher: content-based image search for microscope images

Baek Hwan Cho, Ivan Cao-Berg, Jennifer Ann Bakal & Robert F Murphy

[Affiliations](#) | [Corresponding author](#)

*Nature Methods* **9**, 633–634 (2012) | doi:10.1038/nmeth.2086

Published online 28 June 2012

[Full text](#) [Download PDF](#) [Citation](#) [Reprints](#) [Rights & permissions](#) [Metrics](#)

To the Editor:

Fluorescence microscopy is growing dramatically both in terms of technical capabilities and the volume of images generated. Online repositories have been created to provide public access to images and opportunities for joint research for many scientists<sup>1</sup>. This has reintroduced challenges faced when sequence and structure databases were being established: developing fast and

**Journal home**  
**Current issue**  
**For authors**

**Subscribe**  
**E-alert sign up**  
 **RSS feed**



**nature**  
الطبعة العربية

free Arabic language version  
of *Nature* magazine & regularly  
updated website.

## Selected feature

### Fluorescent proteins and sensors: A practical discussion



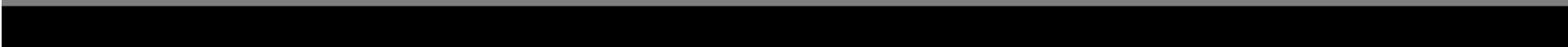
#### Fluorescent proteins and sensors webinar

Robert Campbell, David Piston and Thomas Knöpfel  
Fluorescent proteins are invaluable tools for  
fluorescent microscopy in the life sciences but  
researchers still have practical questions about

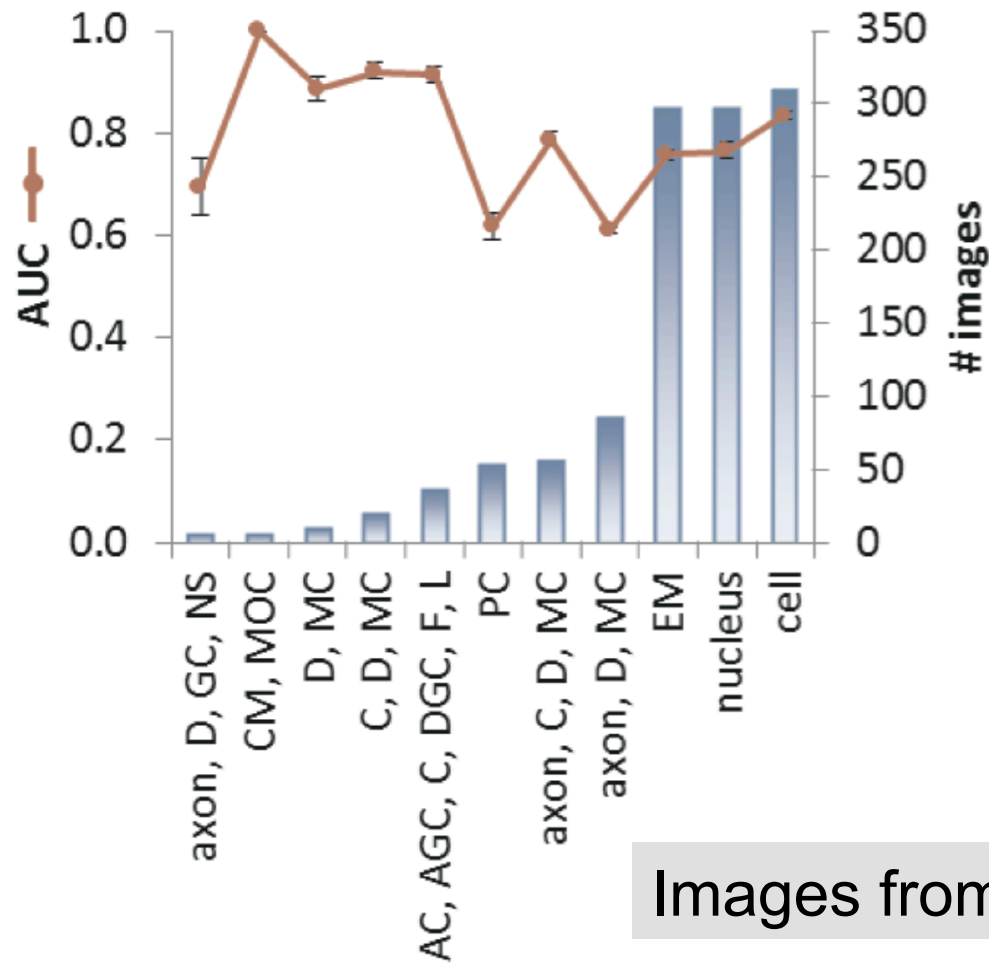




# Testing search effectiveness

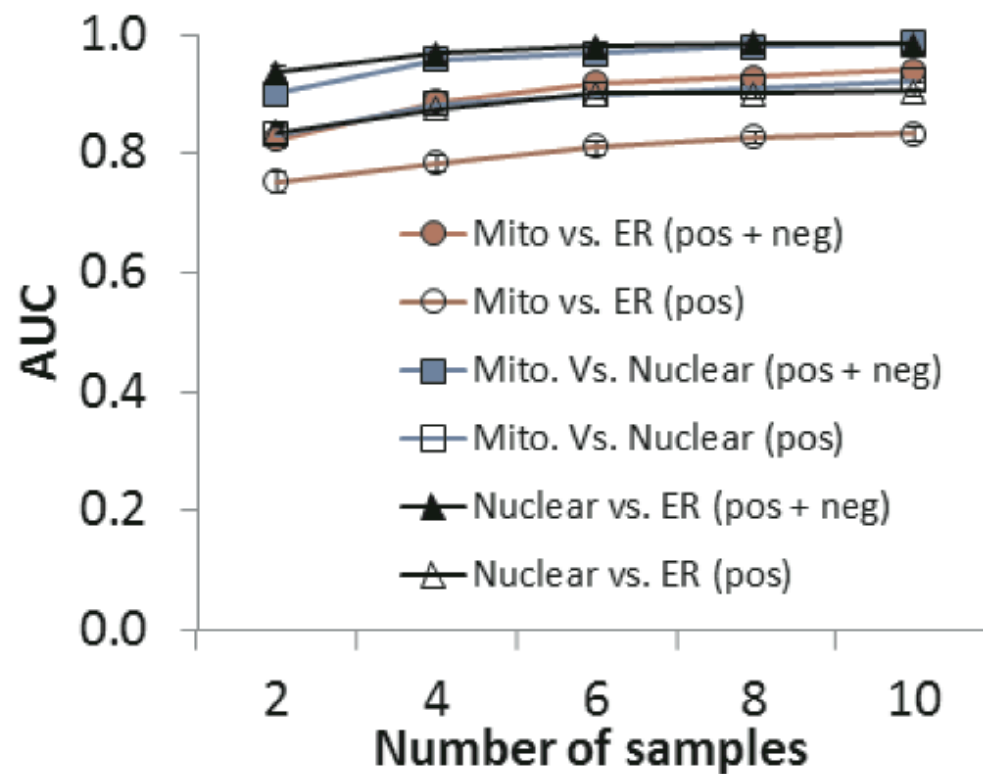
- Combined sets of images of different patterns into database
  - Searched with one or more images of one of the patterns
  - Observed how highly ranked images of that pattern are relative to other patterns (measure area under ROC curve)
- 

# Results for images from diverse image sources



Images from The Cell library

# Results for different patterns from same source

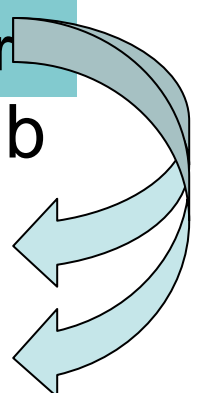


Images from RandTag

# OMERO.searcher

- Open source content-based image database search
- Three ways to use:
  - 1. Server – install on top of your OMERO db
  - 2. External Search – search via web page
  - 3. Local Client – search via Python script

Can search with local images on any computer



Ivan Cao-Berg



Baek Hwan Cho

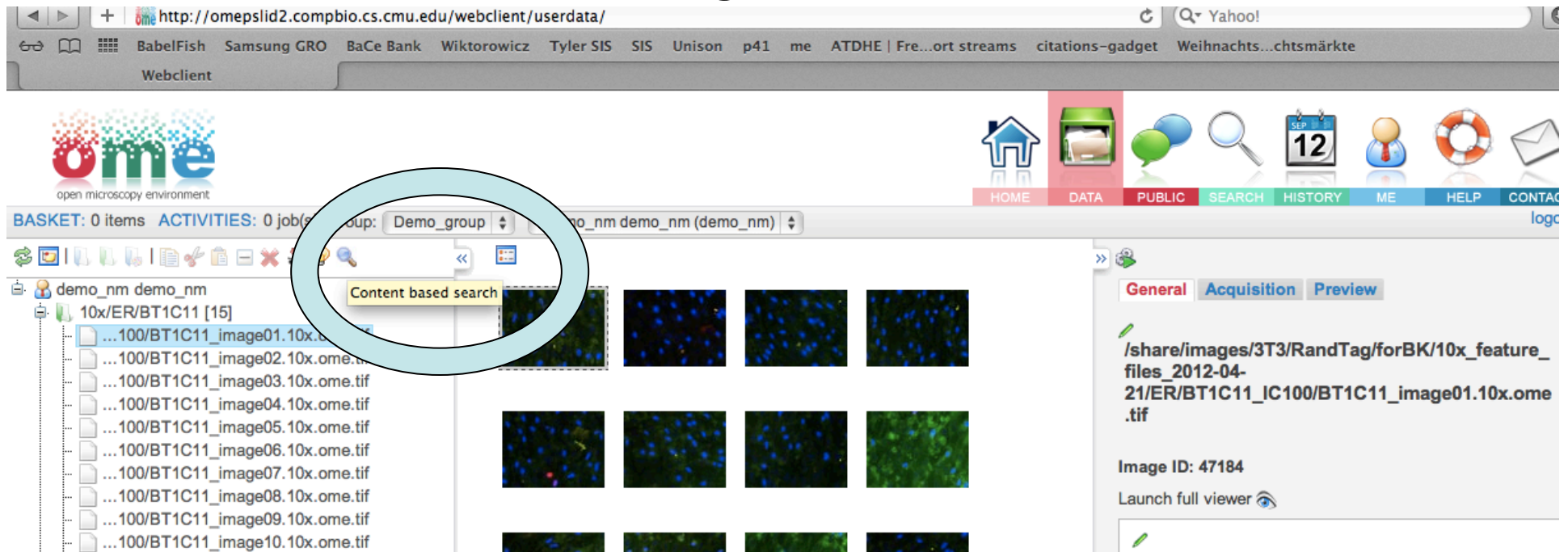


Jenn Bakal

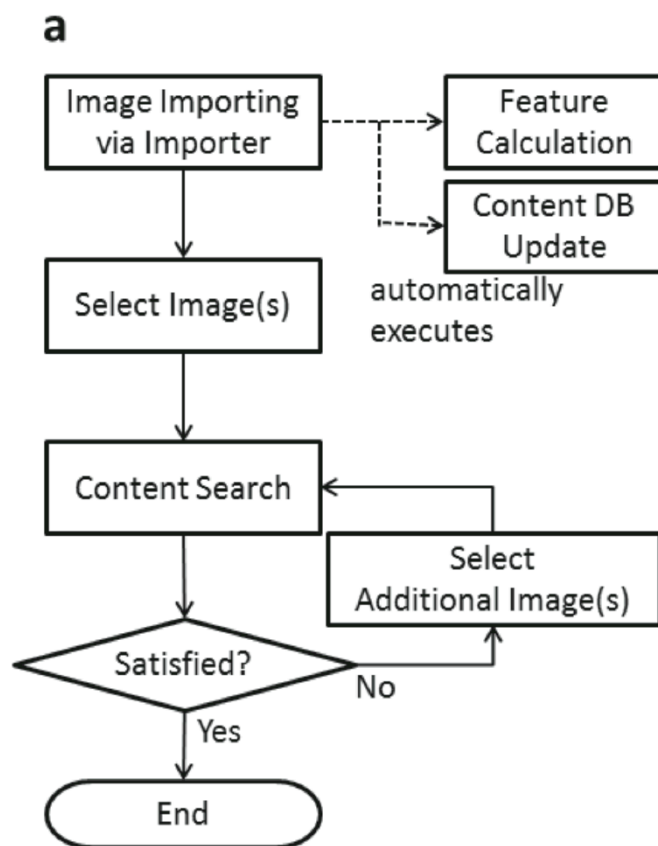


# 1. OMERO.searcher Server

- Install on top of OMERO 4.3
- Installs feature calculation, content search and changes to OMERO.web

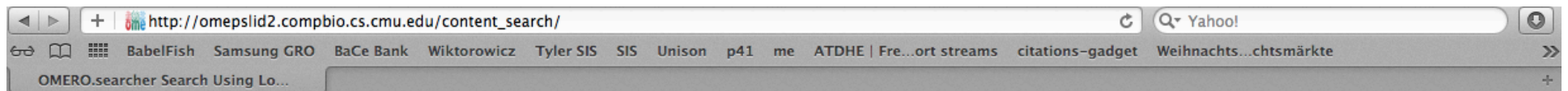


# OMERO.searcher workflow



## 2. External Search page

- Django app installed with Server
- Allows search using external images



### OMERO.searcher 1.1.2

#### Search OMERO database for similar images using local files

This page permits content-based searching of an OMERO database running OMERO.searcher using local image files. The images must be in a format readable by the [Python Imaging Library](#) (which includes most standard image formats like TIFF, PNG, and JPEG), and must contain a single image channel. The two images must also be of the same size and resolution. Images are read by the server, features are calculated and compared to the database, and a list of the most similar images is returned. Images and features are not stored on the server.

Select a primary image file  110509RBT...Z\_001.bmp

Select a reference DNA/nuclear image  110509RBT...Z\_001.bmp

Image scale in microns per pixel

Select number to retrieve

## 2. External Search results



### OMERO.searcher 1.1.2 Results

**Query Image(s)** 110509RBTRBT\_GQ2\_H\_\_6\_T\_001\_ch\_01\_image\_00015\_Z\_001.bmp  
110509RBTRBT\_GQ2\_H\_\_6\_T\_001\_ch\_00\_image\_00015\_Z\_001.bmp

**Content Database** /usr0/local/omero.server/OMERO.server-Beta-  
4.3.3/lib/python/omeroweb/searcher2/contentDB/103\_all\_slf34\_content\_db\_00000000000000000001.pkl

**Original Scale** 0.16125

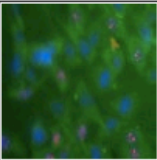
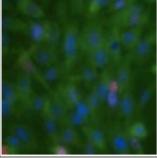

**Comparison Scale** 0.16125

**Feature Set** slf34

**Number to Retrieve** 10

[Return to Content Search](#)

**Results** (if images are not displayed, click on an info link to log into the OMERO database - the username is demo\_nm and the password nature@u0816 - use the back button on your browser to return to this page and then hit refresh)

Thumbnail	Image Data
	<a href="#">Info</a>
	<a href="#">Info</a>
	



## 3. Local Client

- Runs on local computer
- Requires python and packages
- Returns HTML page

Primary  
files

Reference  
files

pixels/  
micron

# to  
return

```
python omero.searcher.py *c1*.bmp *c0*.bmp 0.2 10
```

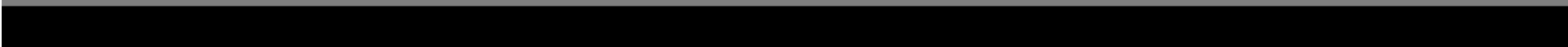


# Local Client version 1.2 released January 8, 2013

- Supports user-specified feature sets
  - Supports non-OMERO databases
- 

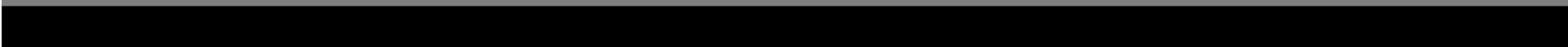


# User-defined feature sets

- Define a feature set as an ordered set of individual features
  - Use python template to produce a function that calculates desired features
- 



# Portable content database

- Contains feature values (for one feature set) for each image in database, with URLs linking to that image
  - Native format is python .pkl file
  - Support for converting to/from .hd5 files
  - Allows embedding of user-defined feature calculation code
- 



# Making a database available for search

- If OMERO database, install OMERO.searcher
- If non-OMERO database, create content DB file containing features and URLs
- In either case, register the content DB so that other users can access, or let people know URL for database in some other manner

# Local Client workflow

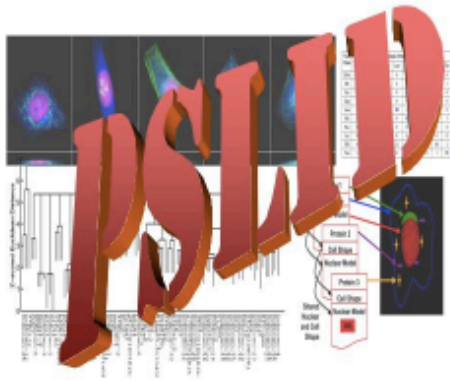
- User specifies database URL, feature set name, query images
- Program
  - looks for content DB for specified DB (and checks if it is latest version)
  - runs feature calculation code on the query images (if features not already calculated)
  - produces HTML file showing results of search

# Publicly searchable databases

- Released
  - Nature Methods demo1
  - **PSLID RandTag Release 2 (new)**
    - <http://pslid.org>
    - 6099 CD-tagged clones, 99955 HTM images, 2777 confocal images, 2937 genomic sequences
    - Identified 116 tagged proteins in 189 clones with high confidence
    - Identified additional 70 tagged proteins in 133 clones with medium confidence

# <http://pslid.org>

## PSLID - Protein Subcellular Location Image Database



*Protein Subcellular Location Image Database*

PSLID is both a web service providing access to collections of images showing subcellular location patterns and an open-source image database software system. There are two main collections: the original [Public database](#) containing various small image sets useful for training and testing various automated analysis approaches, and the [RandTag database](#) containing images of NIH 3T3 clones expressing randomly-tagged proteins.

[Go to PSLID RandTag Release 2](#) (released January 9, 2013)

[Go to the PSLID Public Database Release 4](#) (released August 1, 2008)

[Download datasets contained within PSLID](#)

[Download SLIC feature calculation and pattern analysis software used in PSLID and full PSLID software](#)

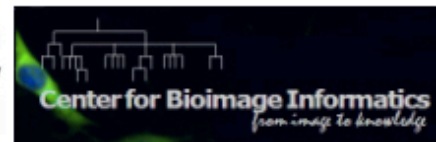
[Go to PSLID RandTag Release 1](#) (released July 1, 2009)



Supported by the National Institute of General Medical Sciences, Grant GM075205

**Carnegie Mellon**

RAY AND STEPHANIE LANE  
Center for Computational Biology



Department  
of Biological  
Sciences





## actin related protein 2/3 complex, subunit 2

- **Gene:** ARPC2\_MOUSE [Ensembl](#)

- **Location from external databases:**

Uniprot	MGI	Locate
<a href="#">cell leading edge:focal adhesion;cell projection;arp2/3 protein complex</a>	<a href="#">golgi apparatus;cytoplasm;cell leading edge:focal adhesion;arp2/3 protein complex;cytoskeleton;cell projection</a>	none found

- **Location from image classification:**

Major(minor)  
cytoplasm (membrane, no\_apparent\_fluorescence); (cytoplasm, membrane, no\_apparent\_fluorescence)

- **Current Curated Location:** none listed  
**Current Proposed Location(s):** none listed

**Proposed Location:**

**email address:**

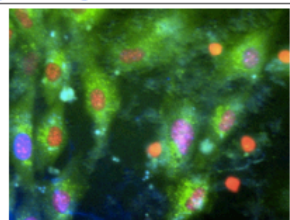
show:  
most typical  
[brightest by dna](#)  
[brightest by protein](#)  
image

### EM1D8\_IC100

[set preview](#) [set details](#)

[sequence information](#)

[gene assignment confidence:](#) Low



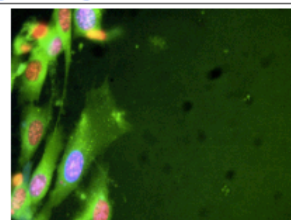
- **Location from image classification:**  
Major class(es): cytoplasm Minor class(es): membrane, no\_apparent\_fluorescence
- **Cell name:** 3T3-CDtag-EM1D8
- **Experiment title:** [3T3-CDtag-EM1](#)
- **Download image files:**

### FX1B4\_IC100

[set preview](#) [set details](#)

[sequence information](#)

[gene assignment confidence:](#) Low



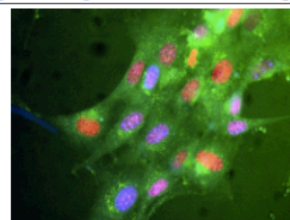
- **Location from image classification:**  
Major class(es): cytoplasm Minor class(es): membrane, no\_apparent\_fluorescence
- **Cell name:** 3T3-CDtag-FX1B4
- **Experiment title:** [3T3-CDtag-FX1](#)
- **Download image files:**

### FX1E9\_IC100

[set preview](#) [set details](#)

[sequence information](#)

[gene assignment confidence:](#) High



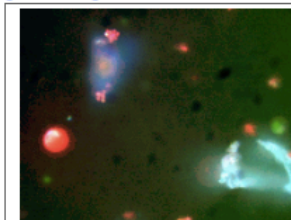
- **Location from image classification:**  
Major class(es): cytoplasm Minor class(es): membrane, no\_apparent\_fluorescence
- **Cell name:** 3T3-CDtag-FX1E9
- **Experiment title:** [3T3-CDtag-FX1](#)
- **Download image files:**

### FX2B3\_IC100

[set preview](#) [set details](#)

[sequence information](#)

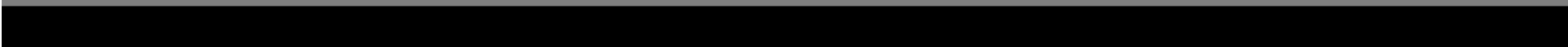
[gene assignment confidence:](#) High



- **Location from image classification:**  
Major class(es): Minor class(es): cytoplasm, membrane, no\_apparent\_fluorescence
- **Cell name:** 3T3-CDtag-FX2B3
- **Experiment title:** [3T3-CDtag-FX2](#)
- **Download image files:**

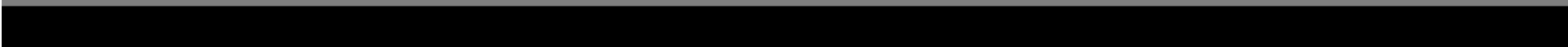


# URL-based searching of PSLID

- Search by target or clone
  - Return HTML or XML
  - Can download images matching a query as a ZIP file of OME-TIFF images
- 



# Planned publicly searchable databases

- The Cell library FM Subset1 (should be released next week)
  - Human Protein Atlas confocal images
- 

# OMERO.searcher plans

- Support for
  - OMERO 4.4 (done?)
  - Search across multiple servers
  - BioFormats for reading images
  - Other feature sets
  - Combining content and context for search
  - Integration into base OMERO system
- Additional OMERO databases available for public search

# PatternUnmixer

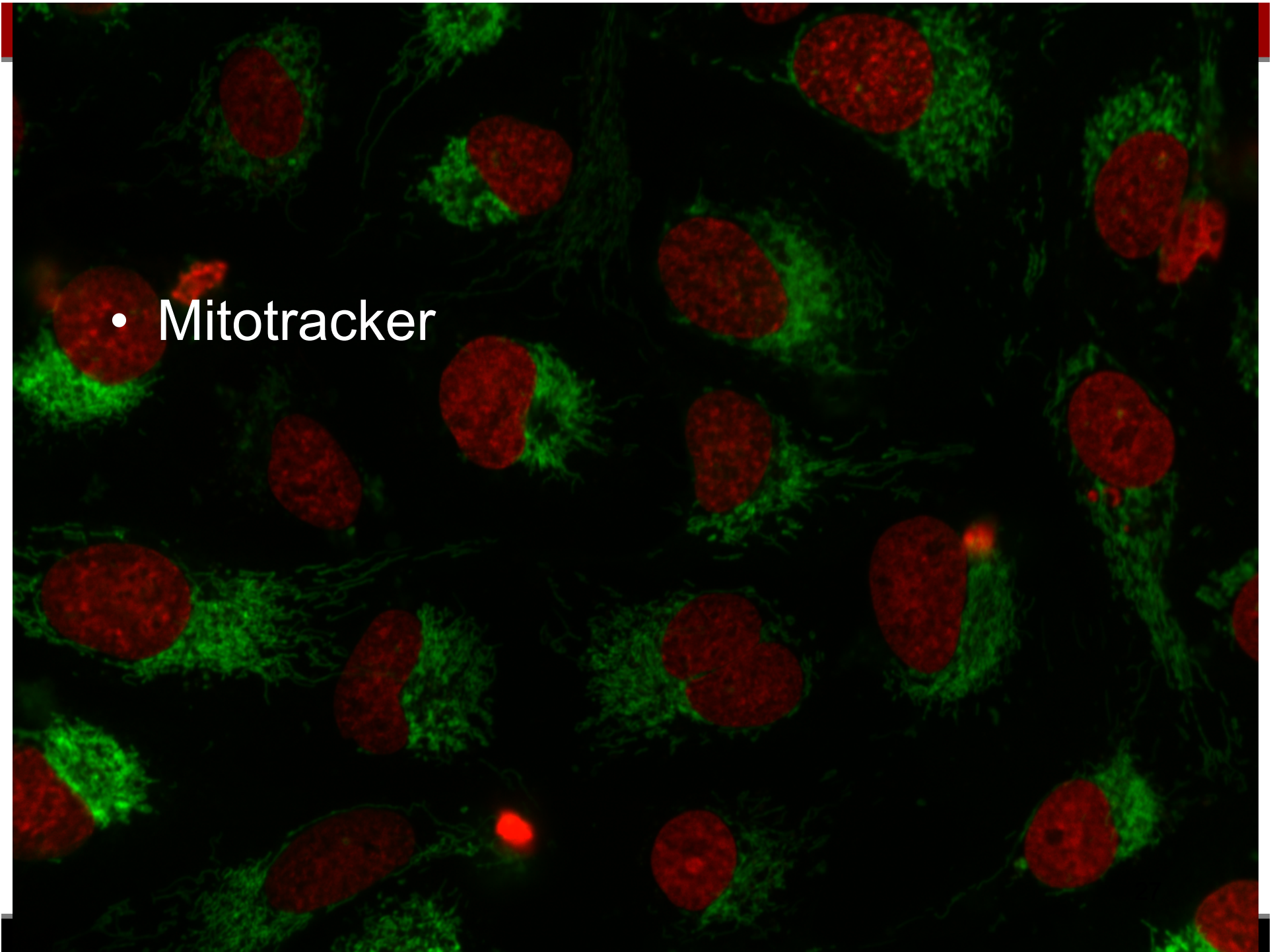
- Performs supervised unmixing of subcellular location
- Versions
  - Matlab source
- Supports training on or unmixing images from local disk or multiple OMERO servers

Tao Peng, Ghislain Bonamy, Estelle  
Glory, Sumit Chanda, Dan Rines  
(Genome Research Institute of  
Novartis Foundation)

- Lysotracker



- Mitotracker

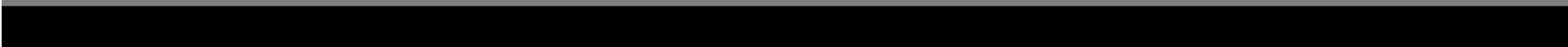




- Mixture of LysoTracker and Mitotracker

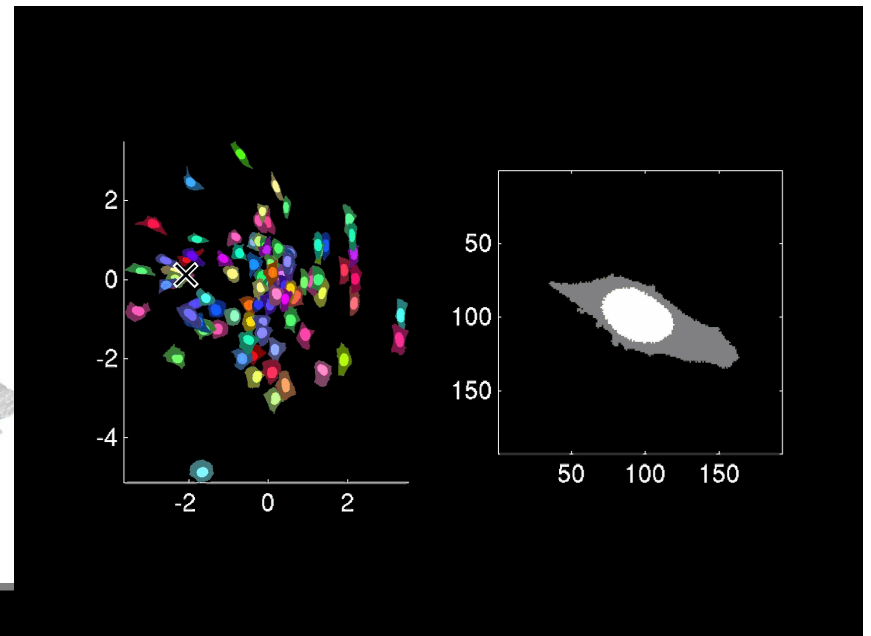
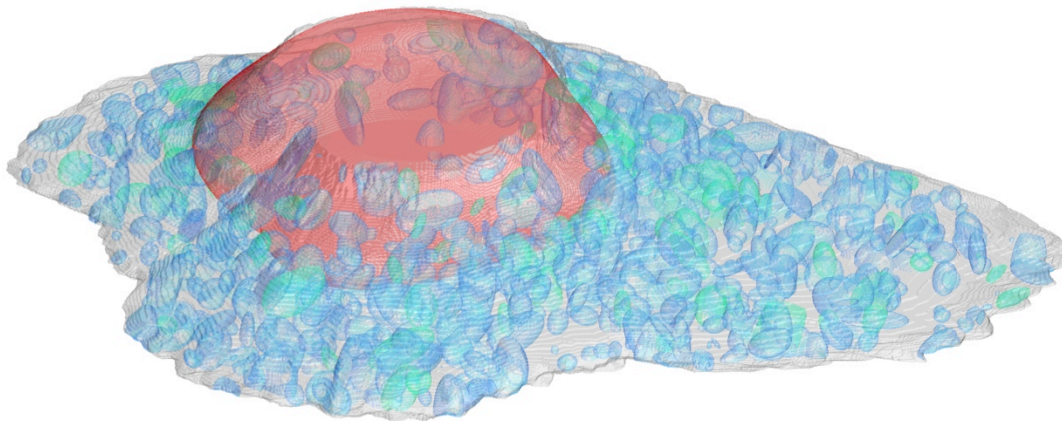


# PatternUnmixer plans

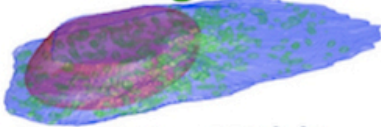
- Django app running on server
    - No requirements on local computer
  - Heatmap displays and hit finders
  - Other local feature detectors
  - Add unsupervised unmixing (have code already)
- 

# CellOrganizer

- Open-source system for learning generative models of cell organization
- Version 1.8 released tomorrow
  - Includes diffeomorphic models for generation of cell and nuclear shapes



# CellOrganizer



Images ↔ Models

[Home](#)  
[People](#)  
[Publications](#)  
[Downloads](#)

April 13, 2012: Integrated 2D/3D [version 1.6](#) released!

The *CellOrganizer* project provides tools for

- learning generative models of cell organization directly from images
- storing and retrieving those models in XML files
- synthesizing cell images (or other representations) from one or more models

Model learning captures variation among cells in a collection of images. Images used for model learning and instances synthesized from models can be two- or three-dimensional static images or movies.

[Version 1.6](#) of *CellOrganizer* can learn models of

- cell shape
- nuclear shape
- chromatin texture
- vesicular organelle size, shape and position
- microtubule distribution.

These models can be *conditional* upon each other. For example, for a given synthesized cell instance, organelle position is dependent upon the cell and nuclear shape of that instance.

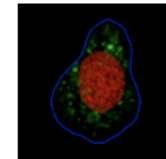
Cell types for which generative models for at least some organelles have been built include human HeLa cells, mouse NIH 3T3 cells, and Arabidopsis protoplasts. Planned projects include mouse T lymphocytes and rat PC12 cells.

RAY AND STEPHANIE LANE  
Center for Computational Biology

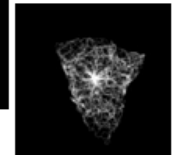
Carnegie  
Mellon  
University

Center for Bioimage Informatics  
*from image to knowledge*

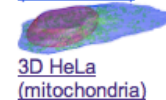
Synthesized Cell Images  
(click to view)



[2D HeLa  
\(endosomes\)](#)



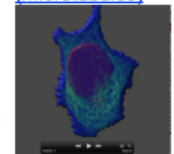
[3D HeLa  
\(microtubules\)](#)



[3D HeLa  
\(mitochondria\)](#)



[3D protoplast  
\(chloroplasts\)](#)



[3D HeLa movie](#)

Support for *CellOrganizer* has been provided by grants GM075205 and GM090033 from the [National Institute of General Medical Sciences](#), by a Forschungspreis from the [Alexander von Humboldt Foundation](#), and by the [School of Life Sciences of the Freiburg Institute for Advanced Studies](#).



Alexander von Humboldt  
Stiftung/Foundation



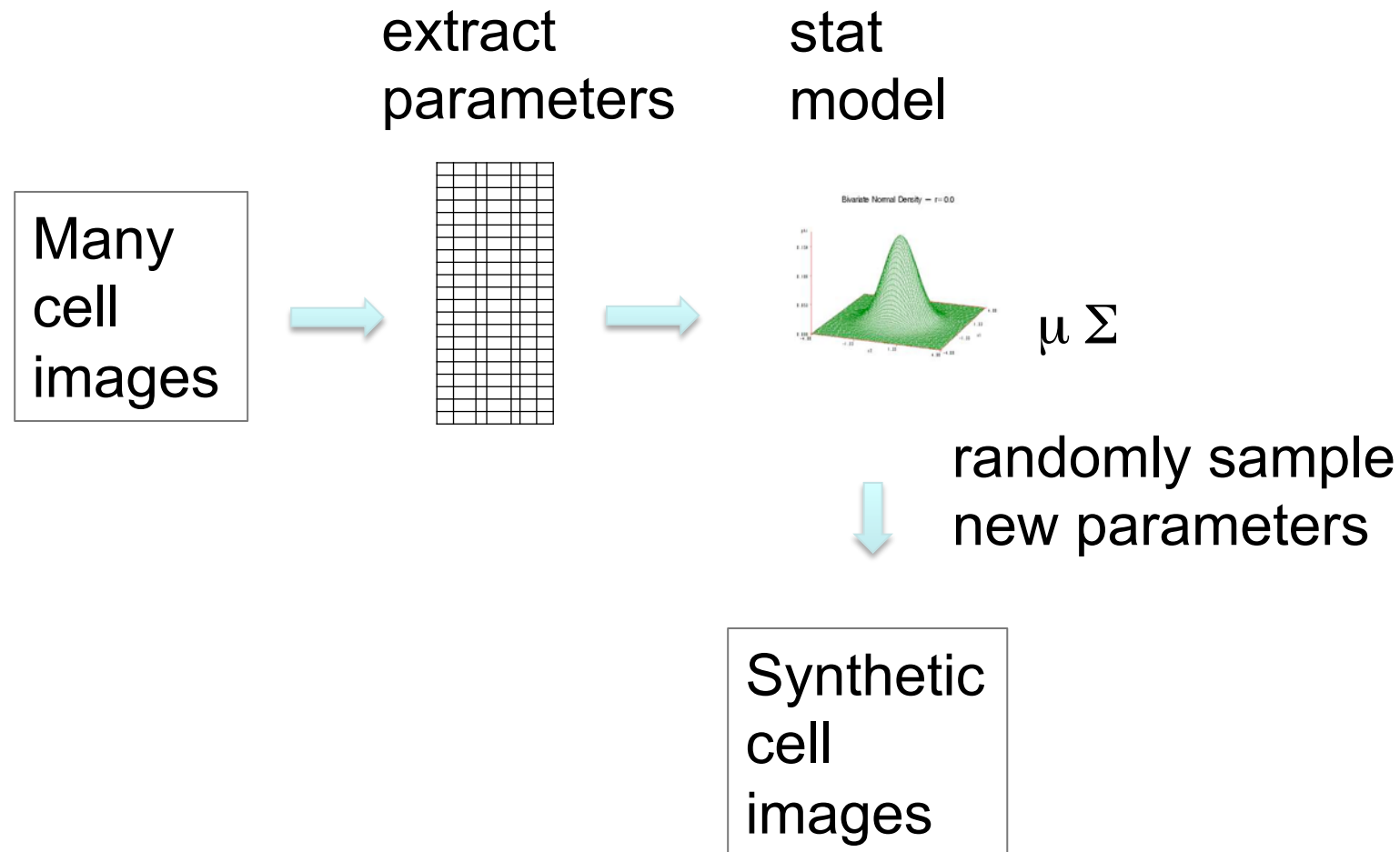
ALBERT-LUDWIGS-  
UNIVERSITÄT FREIBURG



FRIAS  
FREIBURG INSTITUTE  
FOR ADVANCED STUDIES

LIFE SCIENCES - LIFE NET

# Statistical modeling and generation

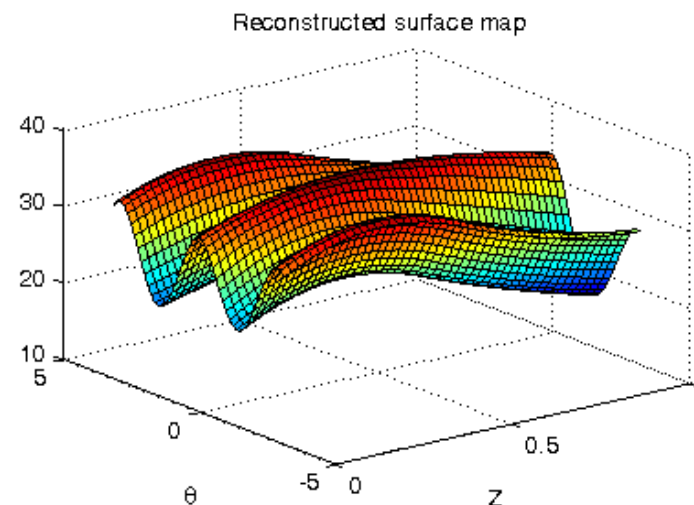
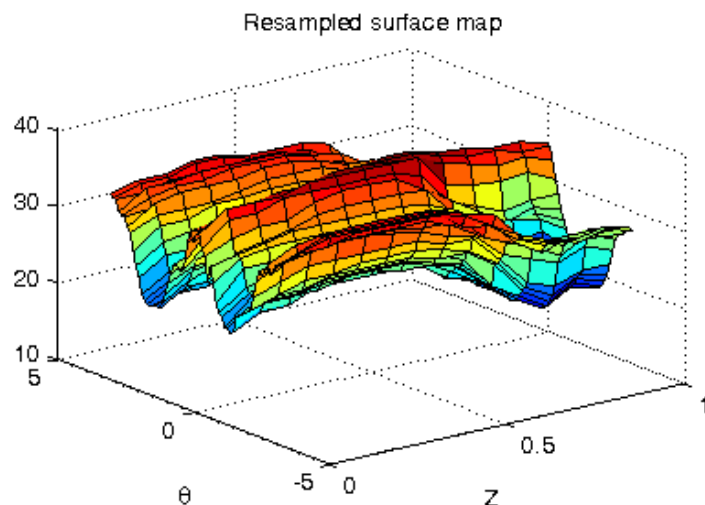
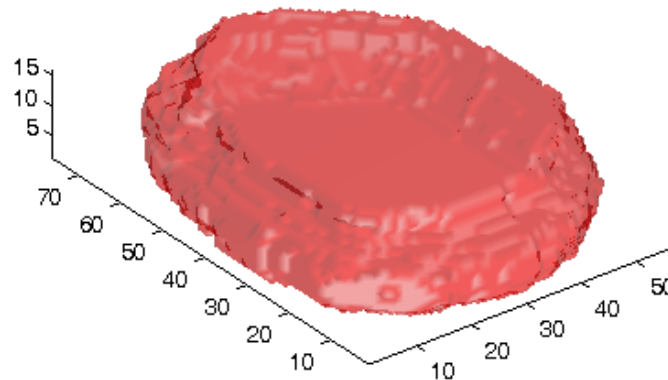




# 3D Nuclear Shape – Cylindrical Spline Surface



Tao Peng



33 parameters  
(32 spline  
coefficients +  
height)

# Diffeomorphic analysis of shape

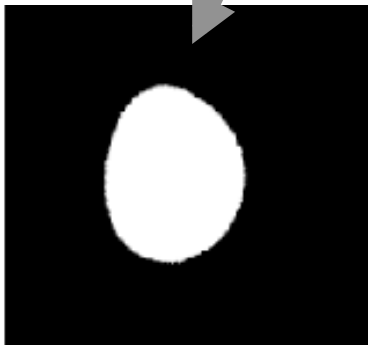
- Sometimes cell or nuclear shapes are irregular
- Can use distance between shapes to characterize shape instead of parameters of model (based on work by Michael Miller and colleagues)

Gustavo  
Rohde



# Morphing one shape into another

Starting shape



0

0.0165

0.0191

0.0194

0.0195

Target shape



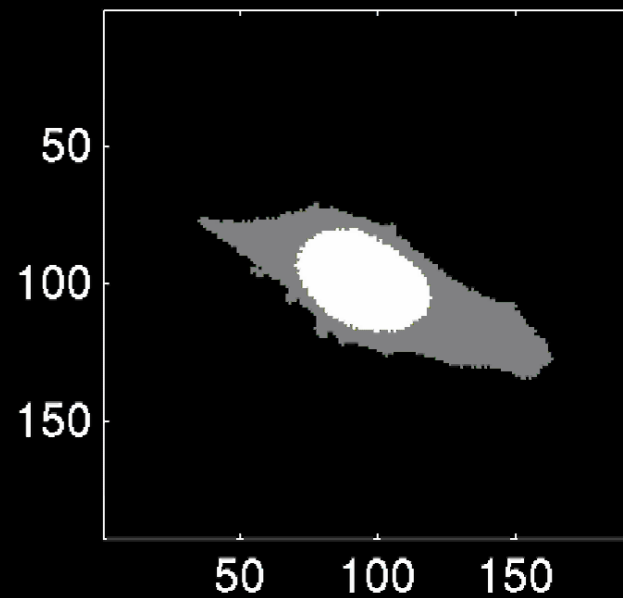
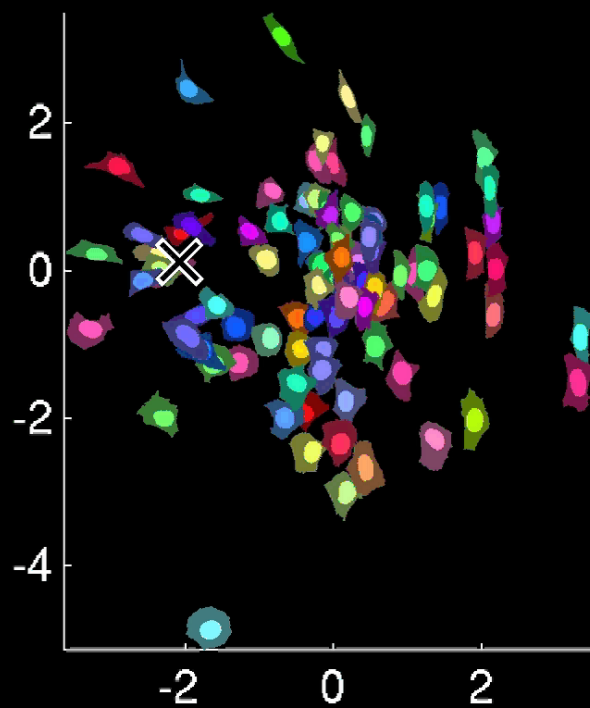
Distance



# Shape space

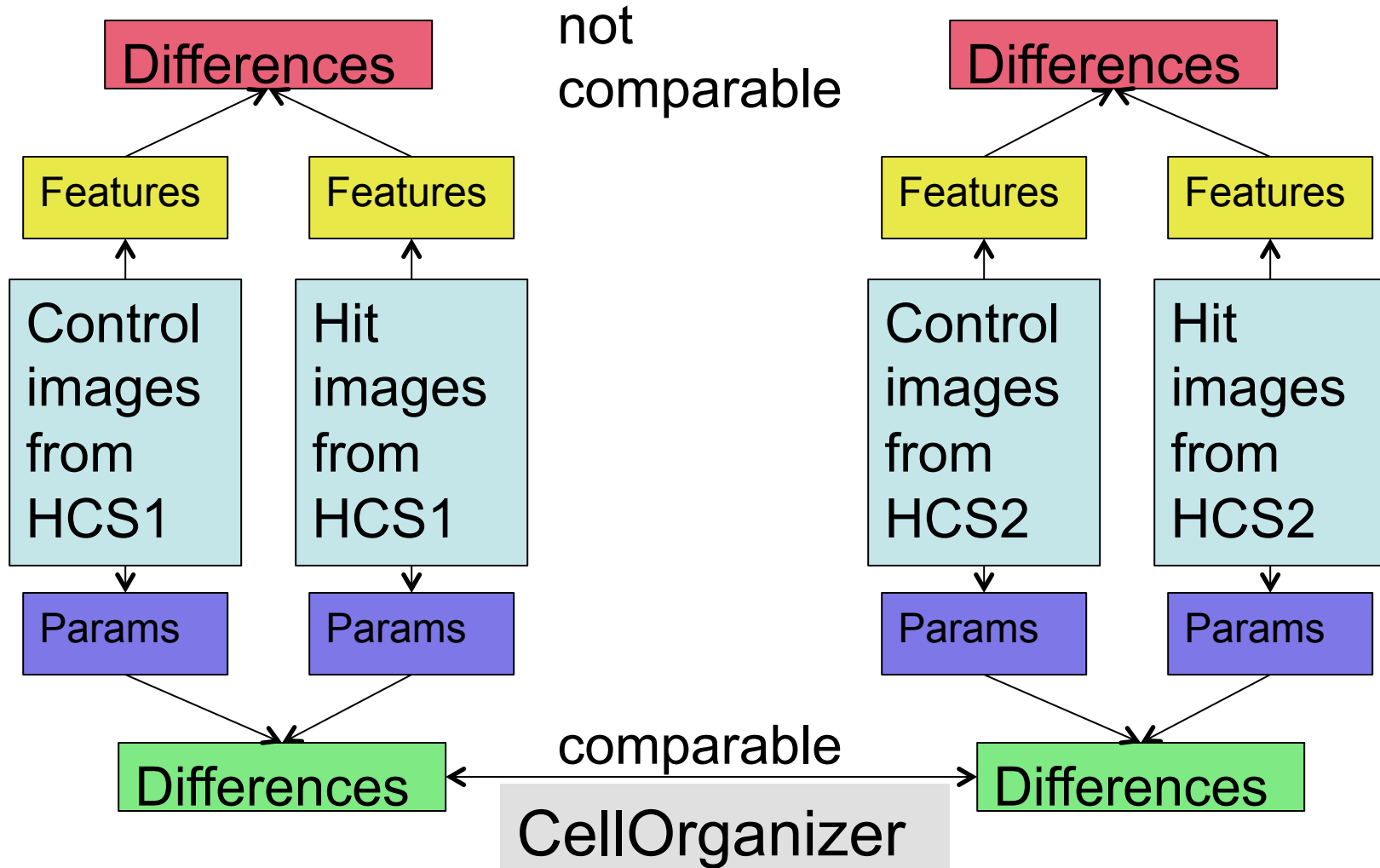
- Can measure distance between all pairs of shapes to construct a “shape space”
- Captures essential aspects of how shapes vary
- Can be applied to nuclei, cells, organelles, etc.
- Can be applied in 2D, 3D, 4D and to more than one component at a time

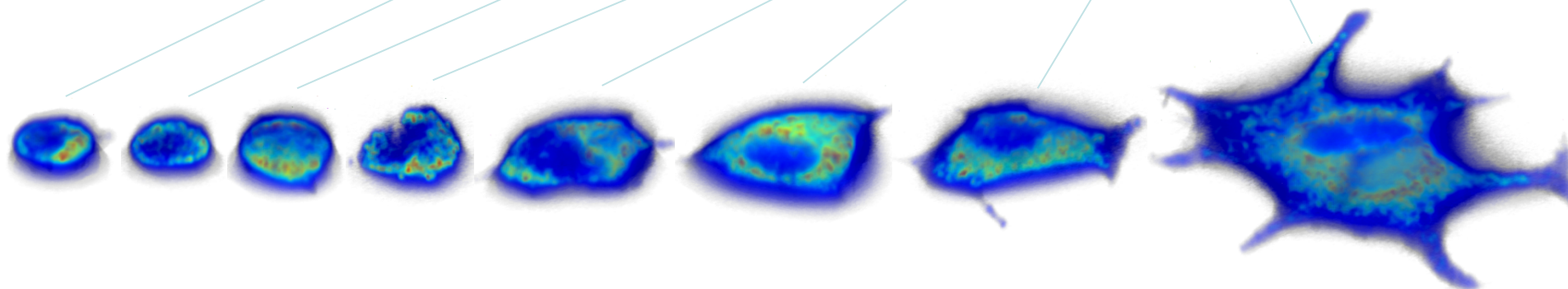
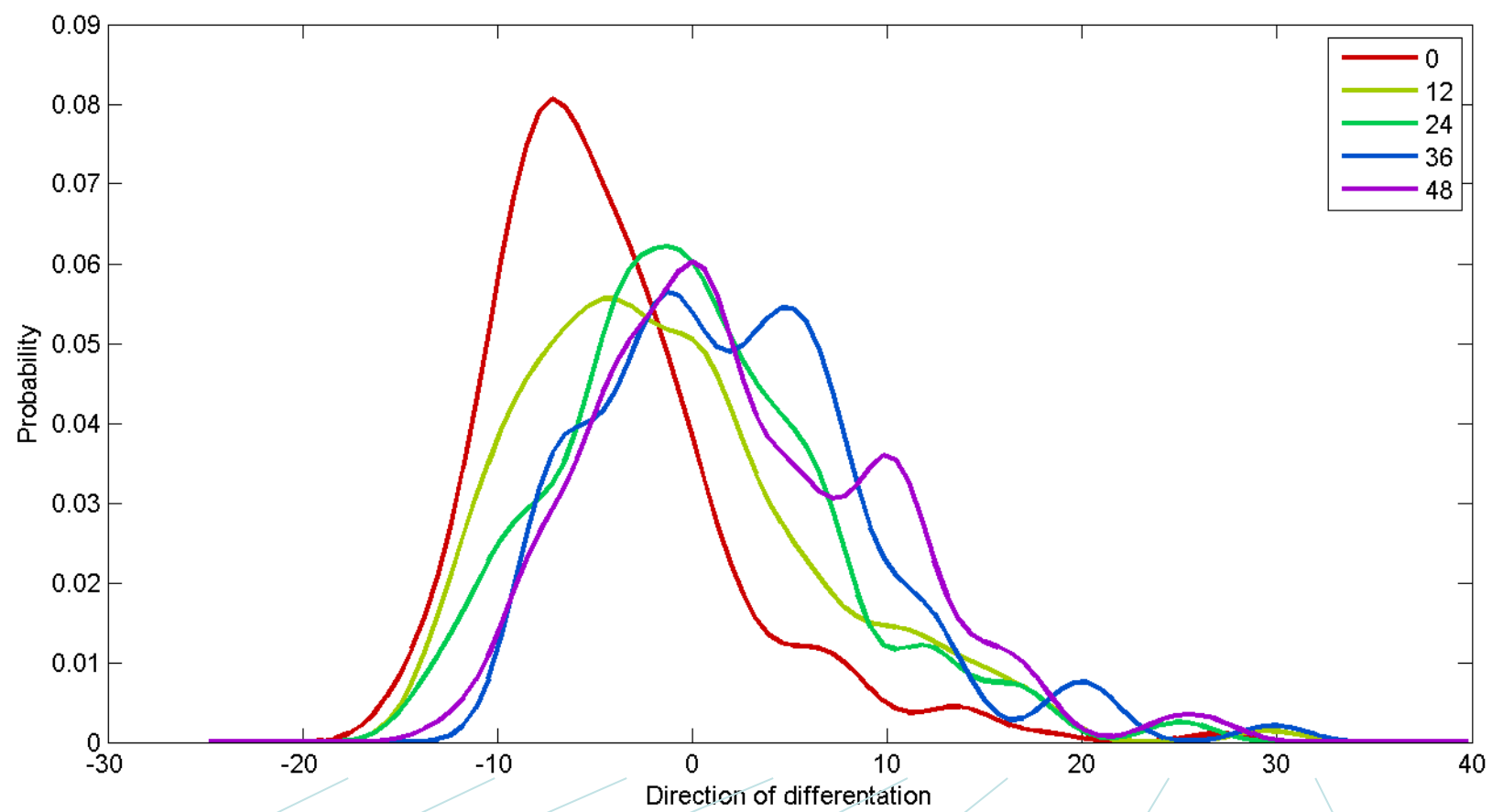
# Cell and nuclear shape space



# CellOrganizer vs. traditional HCS

## Traditional HCS







# A Modest Proposal

- Register the large collection of cellular images within this consortium through generative models
- 